

**Ecography**

**ECOG-03149**

El-Gabbas, A. and Dormann, C. F. 2017. Improved species-occurrence predictions in data-poor regions: using large-scale data and bias correction with down-weighted Poisson regression and Maxent. – Ecography doi: 10.1111/ecog.03149

**Supplementary material**

## Appendix 1: Supplementary figures and tables

**Table A1:** The estimated optimum combination of Maxent's feature classes (FC) and Regularization Multiplier (RM) for each species and bias model type.

Combinations with highest mean testing-AUC on 5-folds spatial-block cross-validation were selected. All the analyses were performed using modified code from the ENMeval package in R (Muscarella *et al.*, 2014). See main text for more information.

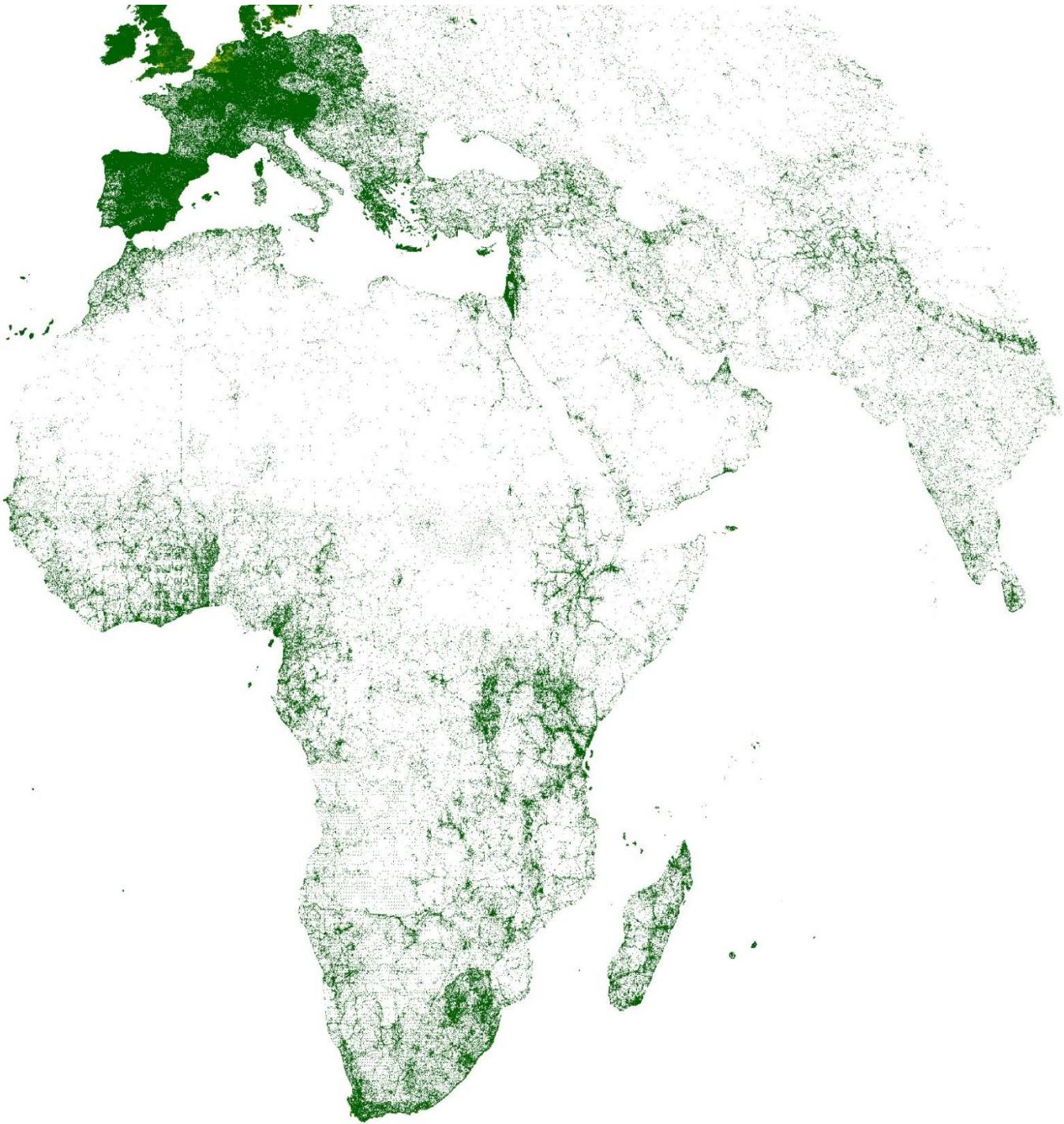
Species	Environment-only		Accessibility		Effort	
	FC <sup>a</sup>	RM <sup>b</sup>	FC	RM	FC	RM
1 <i>Asellia tridens</i>	LQ	0.5	LQ	3	LQHPT	3
2 <i>Barbastella leucomelas</i>	LQ	0.5	LQ	0.5	LQHPT	3.5
3 <i>Eptesicus bottae</i>	LQ	0.5	LQHP	2	LQHP	4
4 <i>Hypsugo ariel</i>	LQHPT	2	LQHP	2	L	1
5 <i>Nycterus thebaica</i>	LQ	4	LQ	3.5	LQ	4
6 <i>Nycticeinops schlieffeni</i>	LQ	1	LQ	0.5	LQ	1.5
7 <i>Otonycteris hemprichii</i>	LQ	0.5	LQ	1	LQHPT	0.5
8 <i>Pipistrellus deserti</i>	L	2.5	LQ	0.5	L	3
9 <i>Pipistrellus kuhlii</i>	LQ	0.5	LQ	0.5	LQ	0.5
10 <i>Pipistrellus rueppellii</i>	LQ	4	LQ	0.5	LQ	4
11 <i>Plecotus christii</i>	LQ	0.5	LQHPT	0.5	LQHPT	0.5
12 <i>Rhinolophus clivosus</i>	LQHP	2.5	LQHPT	2	LQHPT	2.5
13 <i>Rhinolophus hipposideros</i>	LQ	0.5	LQ	0.5	LQ	0.5
14 <i>Rhinolophus mehelyi</i>	LQ	0.5	LQ	0.5	LQ	2.5
15 <i>Rhinopoma cystops</i>	LQ	1	LQ	1.5	LQHPT	1
16 <i>Rhinopoma microphyllum</i>	LQ	1.5	LQ	2	LQHPT	4
17 <i>Rousettus aegyptiacus</i>	LQHPT	2.5	LQ	3	LQ	4
18 <i>Tadarida aegyptiaca</i>	LQ	3	LQ	4	LQ	3
19 <i>Tadarida teniotis</i>	LQ	0.5	LQ	0.5	LQ	0.5
20 <i>Taphozous nudiventris</i>	LQ	1	L	2	LQHPT	2.5
21 <i>Taphozous perforatus</i>	LQ	1.5	LQ	1	LQ	1.5

<sup>a</sup> Feature classes (FC): **L** linear; **Q** quadratic; **H** hinge; **P** product; and **T** threshold. For more details see Elith *et al.* 2011.

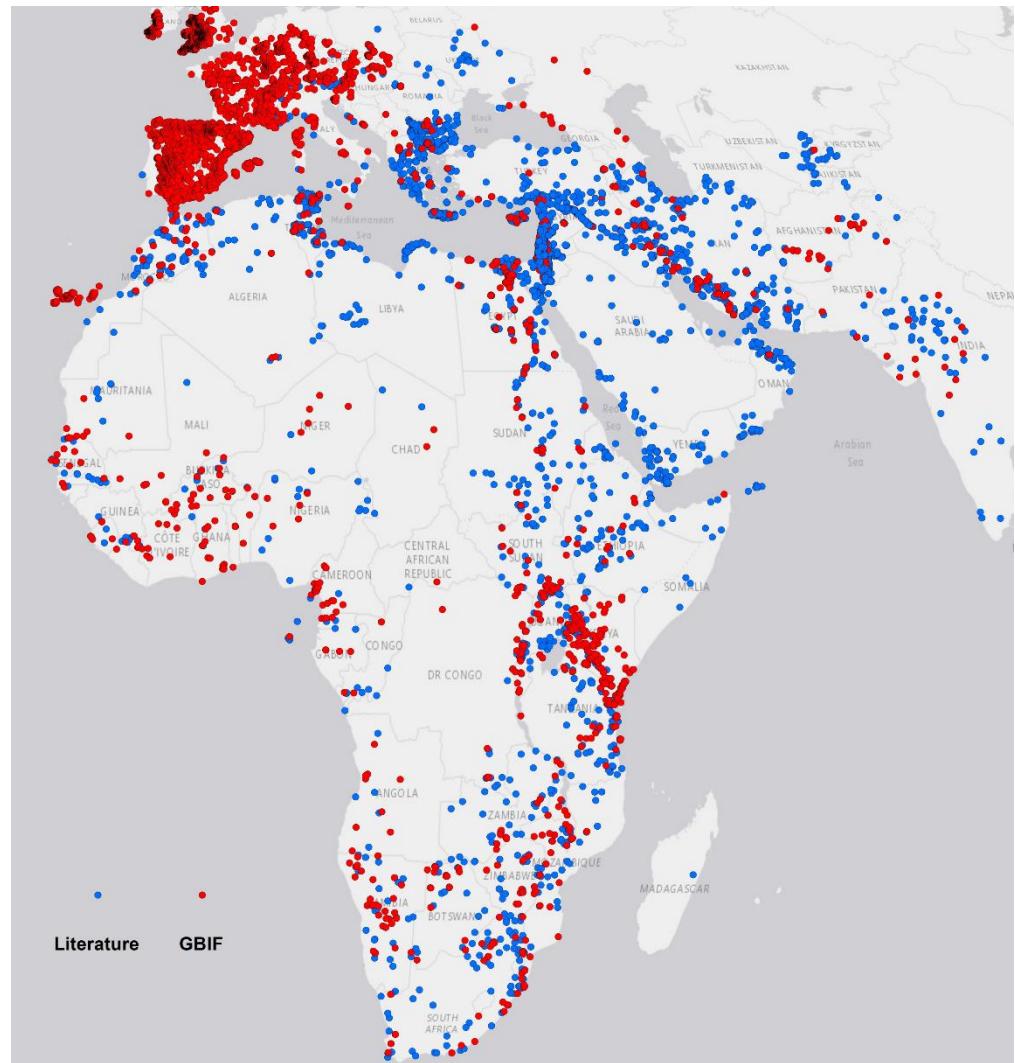
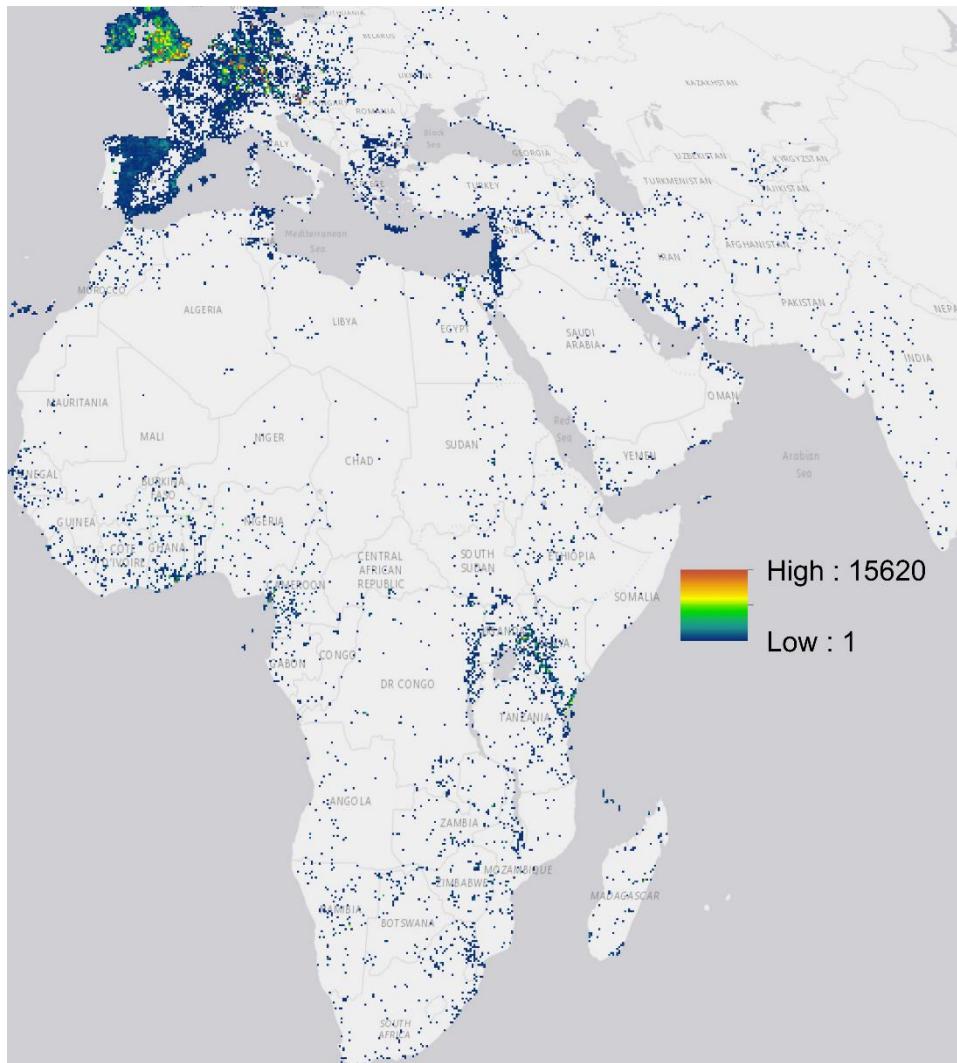
<sup>b</sup> RM is a global multiplier to all individual regularization values; for more details see the main text and Muscarella *et al.* (2014).

**Table A2:** Estimated best  $\alpha$ -values to run the elastic-net models (DWPR approach) for different species and bias models. For details, see main text.

	<b>Species</b>	<b>Environment-only</b>	<b>Accessibility</b>	<b>Efforts</b>
1	<i>Asellia tridens</i>	1 (LASSO)	1 (LASSO)	0 (RIDGE)
2	<i>Barbastella leucomelas</i>	1 (LASSO)	0.9	0.7
3	<i>Eptesicus bottae</i>	0.3	0.8	0.1
4	<i>Hypsugo ariel</i>	1 (LASSO)	1 (LASSO)	0.2
5	<i>Nycterus thebaica</i>	1 (LASSO)	0.1	0.5
6	<i>Nycticeinops schlieffeni</i>	1 (LASSO)	1 (LASSO)	0.6
7	<i>Otonycteris hemprichii</i>	1 (LASSO)	1 (LASSO)	0.2
8	<i>Pipistrellus deserti</i>	0 (RIDGE)	0.8	0 (RIDGE)
9	<i>Pipistrellus kuhlii</i>	0.8	0.2	0.9
10	<i>Pipistrellus rueppellii</i>	0 (RIDGE)	0 (RIDGE)	0 (RIDGE)
11	<i>Plecotus christii</i>	0.6	0.1	0.5
12	<i>Rhinolophus clivosus</i>	0.1	0.1	0.2
13	<i>Rhinolophus hipposideros</i>	0.4	0.4	0.3
14	<i>Rhinolophus mehelyi</i>	1 (LASSO)	0.1	0.6
15	<i>Rhinopoma cystops</i>	1 (LASSO)	1 (LASSO)	0.6
16	<i>Rhinopoma microphyllum</i>	0.4	0.9	0.9
17	<i>Rousettus aegyptiacus</i>	0.7	1 (LASSO)	0.9
18	<i>Tadarida aegyptiaca</i>	1 (LASSO)	1 (LASSO)	0.9
19	<i>Tadarida teniotis</i>	0.5	0.4	0.7
20	<i>Taphozous nudiventris</i>	0.9	1 (LASSO)	0.8
21	<i>Taphozous perforatus</i>	0.1	1 (LASSO)	0.5



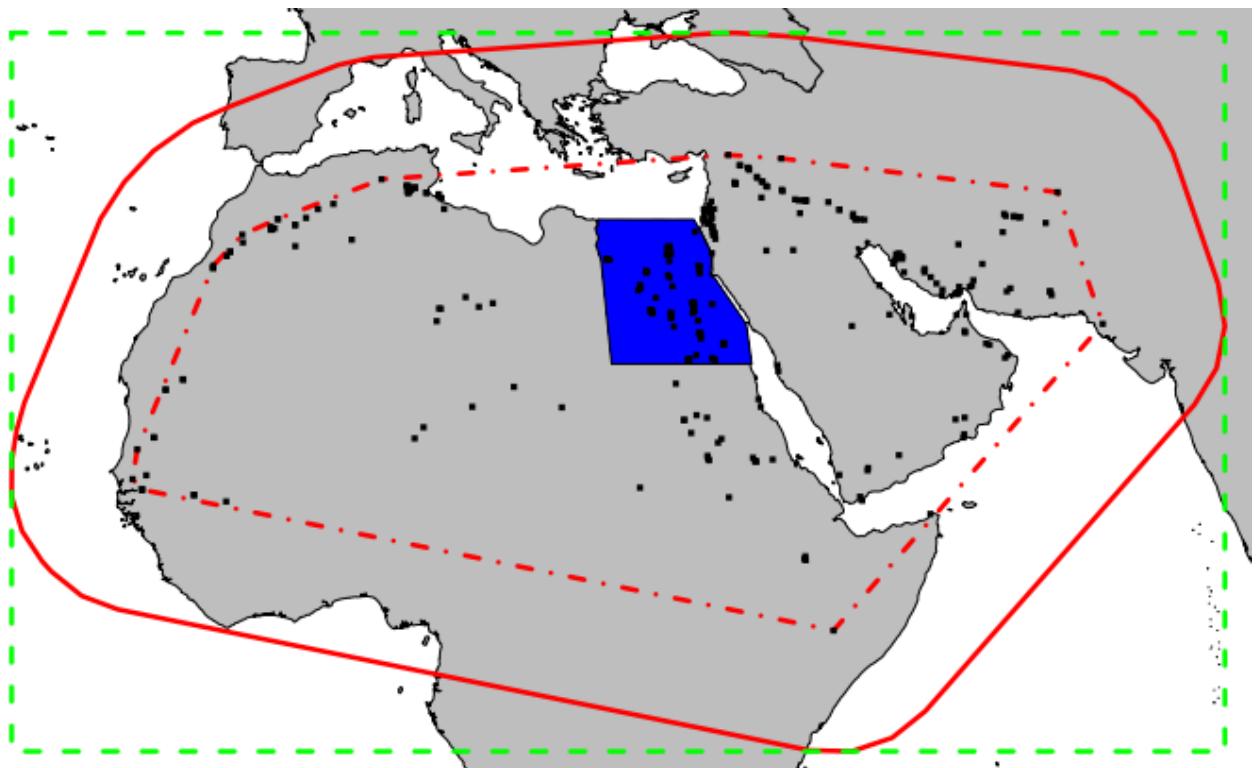
**Fig. A1:** Map of all non-marine GBIF-records. There are clear signs of sampling bias inherited in the GBIF database, disallowing the direct use of these records to run SDMs, unless action is taken to correct for sampling bias. GBIF-records are biased mainly towards Western Europe, with sparse locations across Africa, Asia, and Eastern Europe. Areas shown in white represent pixels without any records (the majority of North and central Africa, Arabia, and eastern Europe). Clearly erroneous records were excluded before plotting this map.



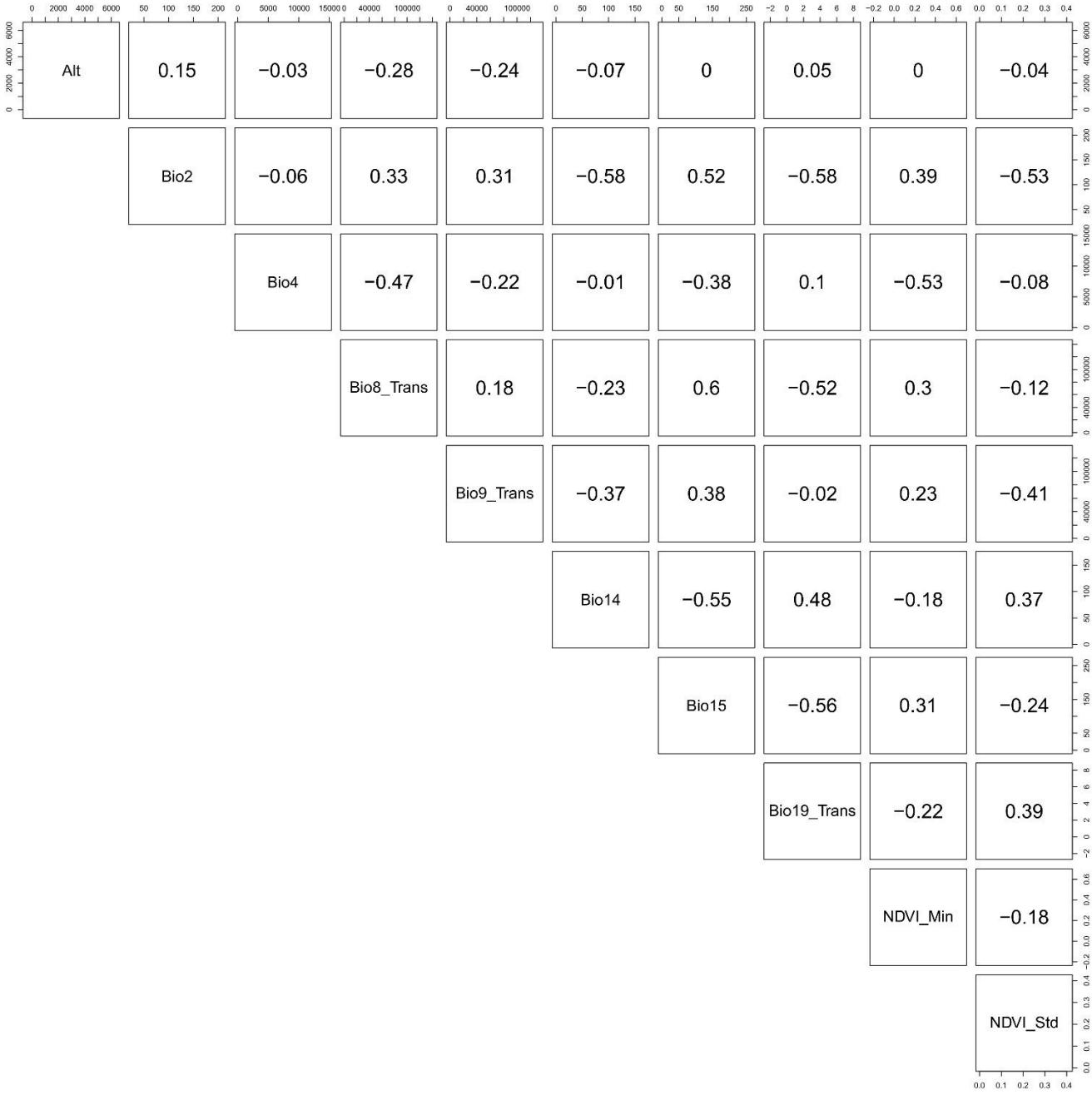
**Fig. A2:** Left: Map of all GBIF bat records (accessed date: April 2015) per a grid of 20 × 20 km<sup>2</sup>.

Right: All records available of the study species, collected either from the literature (blue points) or GBIF (red points).

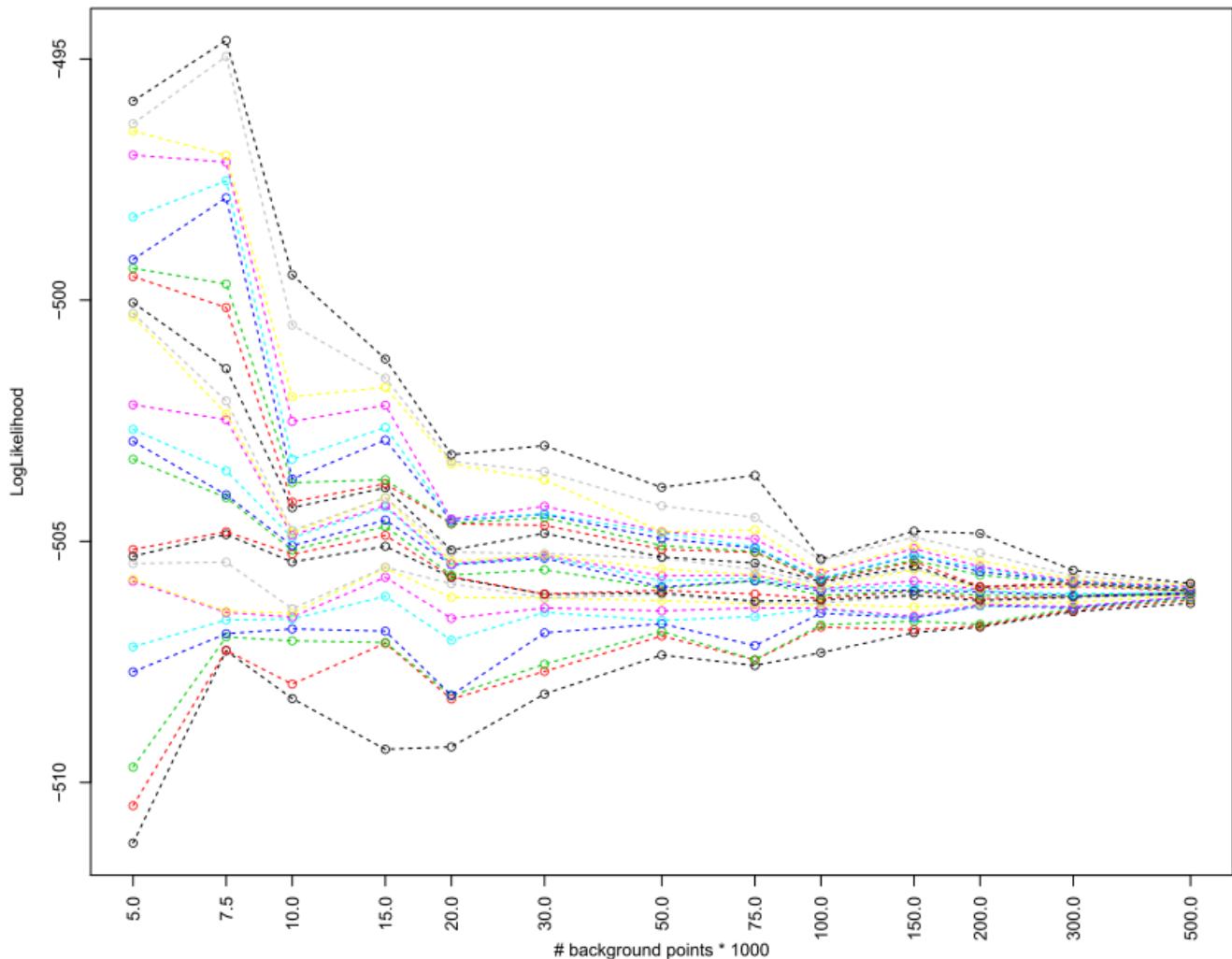
Both maps show large collection gaps.



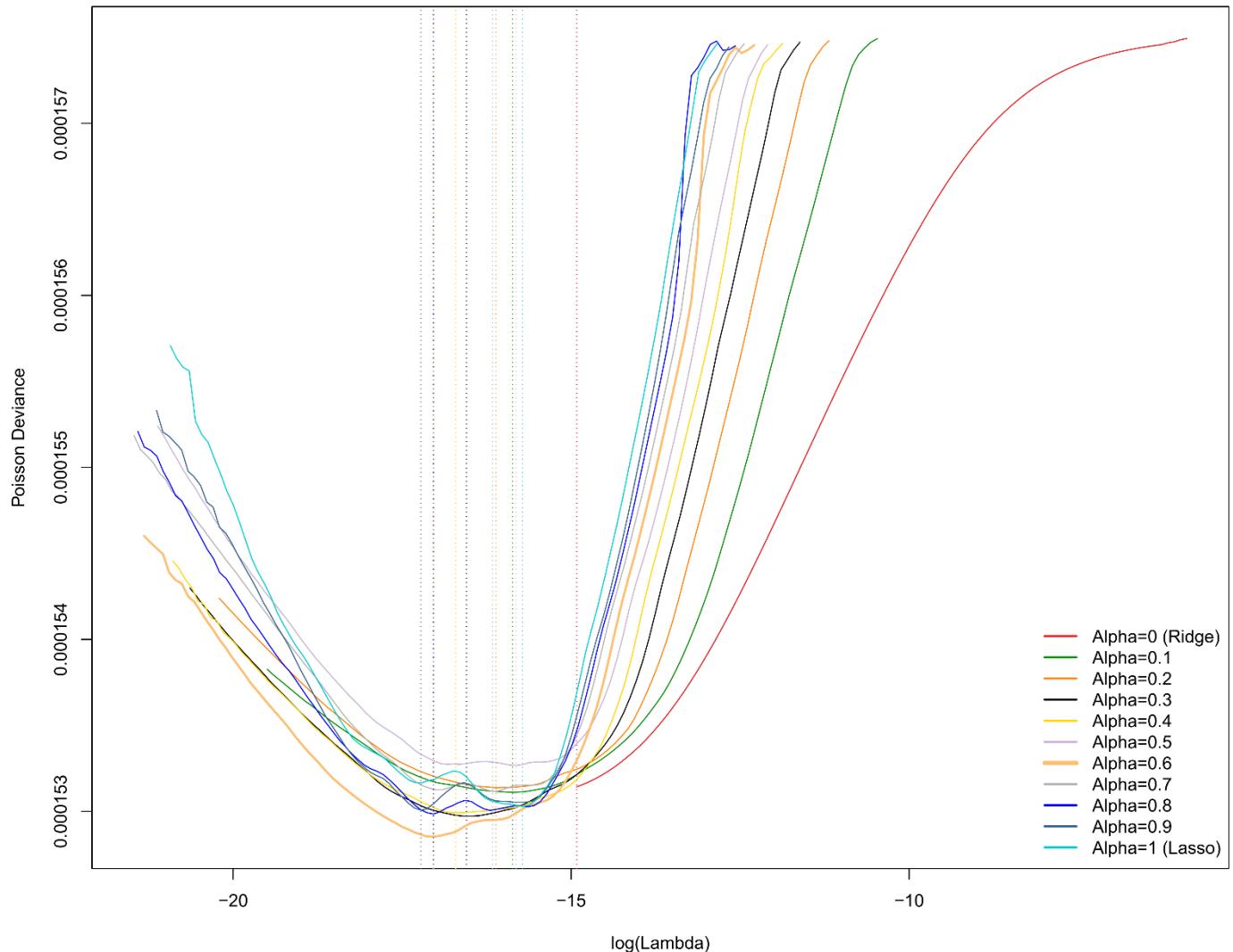
**Fig. A3: Per-species study area:** An example of how the 'per-species' study area was determined (here for *Asellia tridens*). The dashed red line indicates the species extent of occurrence (minimum convex polygon); the solid red line shows the buffer used (1000 km), and the dashed green line indicates the study area used.



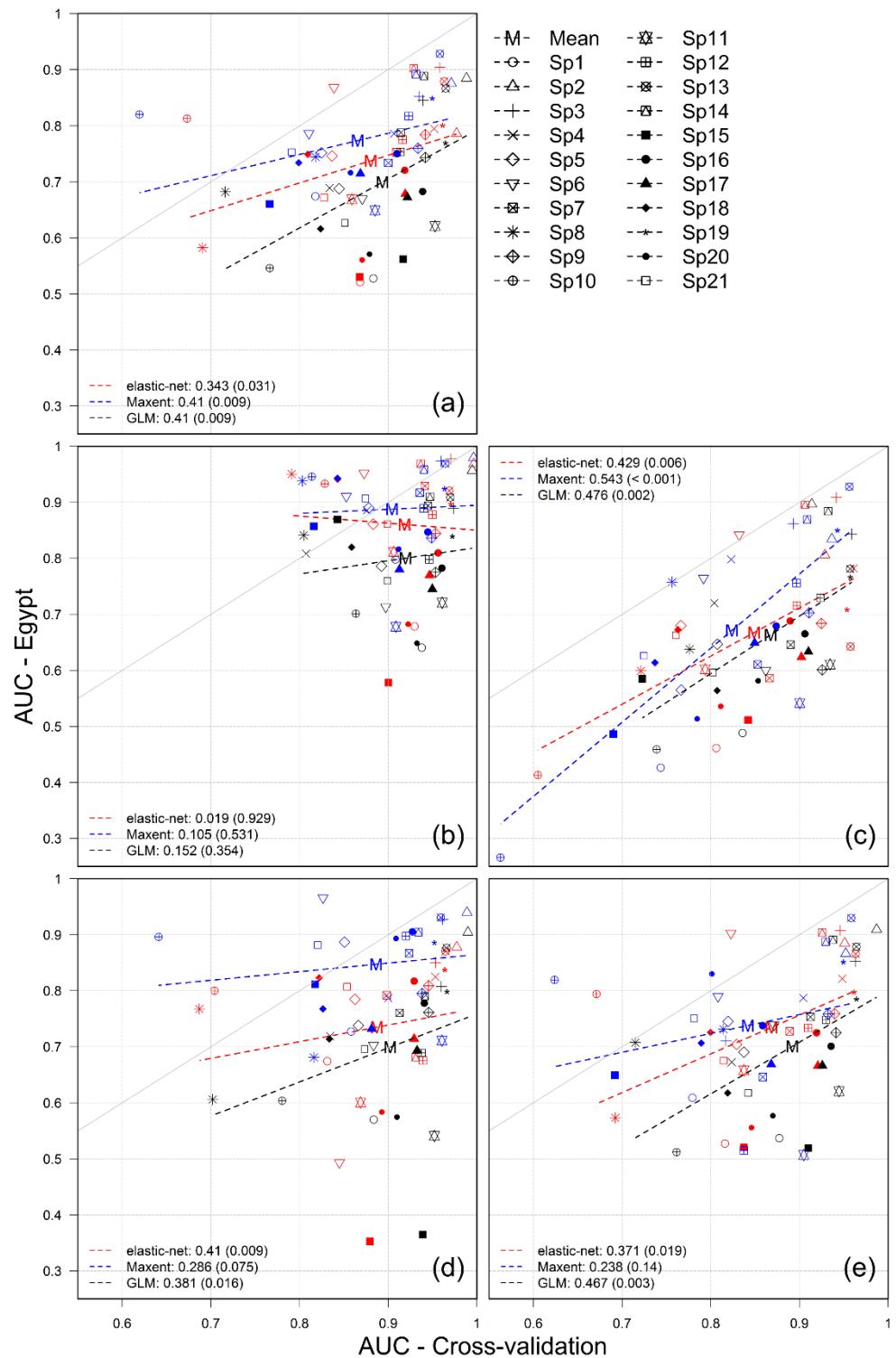
**Fig. A4: Variables selection:** Pearson correlation coefficient between each pair of the final covariates; see Appendix 3 for more information.



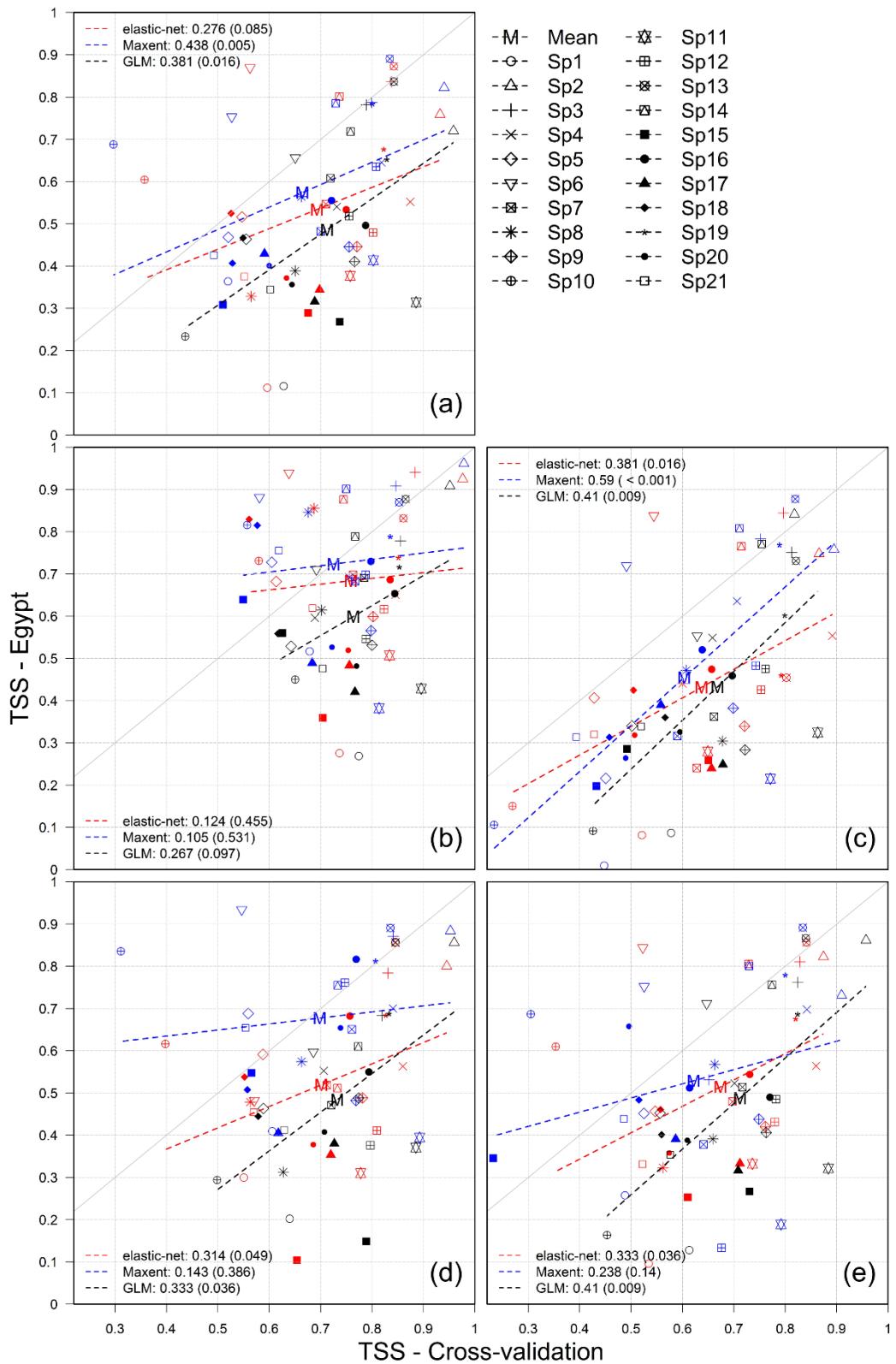
**Fig A5: Per-species number of backgrounds:** Objectively determining the adequate number of background ‘quadrature’ points to be used in GLM and elastic-net models (see main text and Renner *et al.*, 2015). DWPR-GLM models (for each species, here: *Asellia tridens*) were repeated 25 times (each time, with a different random sample from the background) with progressively increase the number of background points (from 5000 to 500,000); and for each model, the log-likelihood value is calculated and plotted. The number of backgrounds at which the likelihood converges (i.e. no much benefit of using a higher number of backgrounds) is, visually, selected to run the final models. Here, no much improvement in the log-likelihood beyond 300,000 background points (out of 557,800 total available background points) and so we used it for this species.



**Fig. A6:** Determining the best  $\alpha$ -value to run the elastic-net model for *Nycticeinops schlieffeni* (effort model). Eleven  $\alpha$ -values ranging from zero (ridge) to one (lasso), with an increment of 0.1, were used to run 11 models on cross-validation. For each value of  $\alpha$ , glmnet ran many models on a range of  $\lambda$ -values (estimated from the data; x-axis) and measure the mean cross-validation error (cvm) (here: Poisson deviance; y-axis). The  $\lambda$ -value with the lowest cvm is used further while predicting (vertical dotted lines, with colours correspond to the  $\alpha$ -value used). Here, an  $\alpha$ -value of 0.6 is shown to have the lowest cvm and so used further. For more details, see Table A2 and the main text.



**Fig. A7:** Kendall's correlation between (per-species and modelling algorithm) mean AUC evaluated on spatial-block cross-validation and independent evaluation in Egypt. Each point represents the mean of 100 AUC values, with different symbols for each species. Colours represent different modelling algorithm used. 'M' indicates the overall mean for each modelling algorithm. Panel (a) shows the correlation for the environment-only models (no bias correction). Panels (b-c) represent the accessibility model, without and with correcting for sampling bias, respectively. Panels (d-e) represent the effort model, without and with correcting for sampling bias, respectively. Kendall's correlation coefficients (and their p-values) are reported in each block. Grey solid line represents the equality line. AUC performed on cross-validation were, on average, higher than AUC in Egypt. However, there is moderate correlation between bias-free evaluations on either scale.



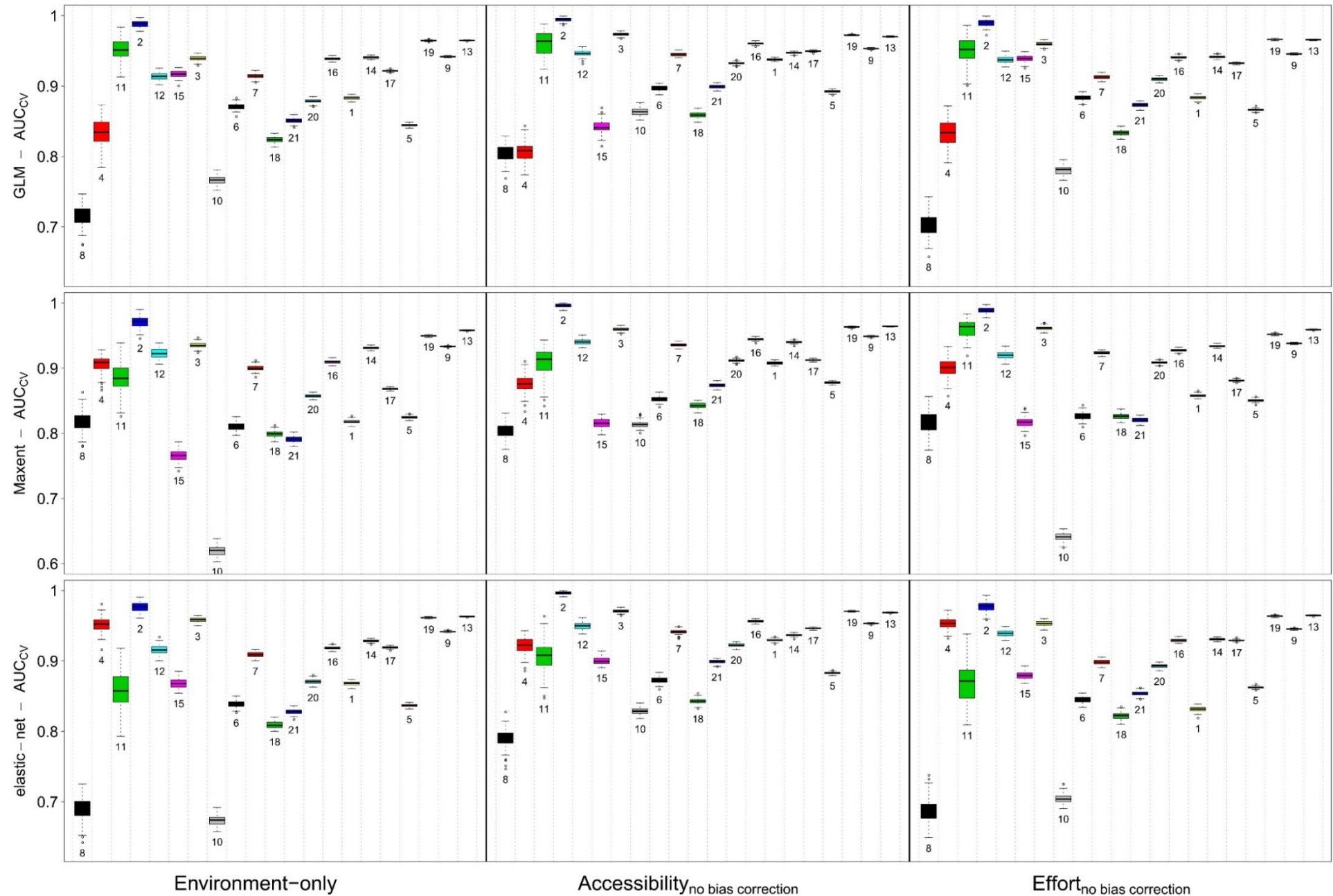
**Fig. A8:** Equivalent results to Fig. A7, for TSS evaluation.

**Fig A9:** Boxplots of the raw cross-validation evaluation (100 values) without (modelling evaluation; a and c) and with (bias-free evaluation; b and d) correction for sampling bias; either using AUC (a - b) or TSS (c - d); see Appendix 6 for more information. Horizontal panels show results for different modelling algorithm, while the vertical panels show different bias models.

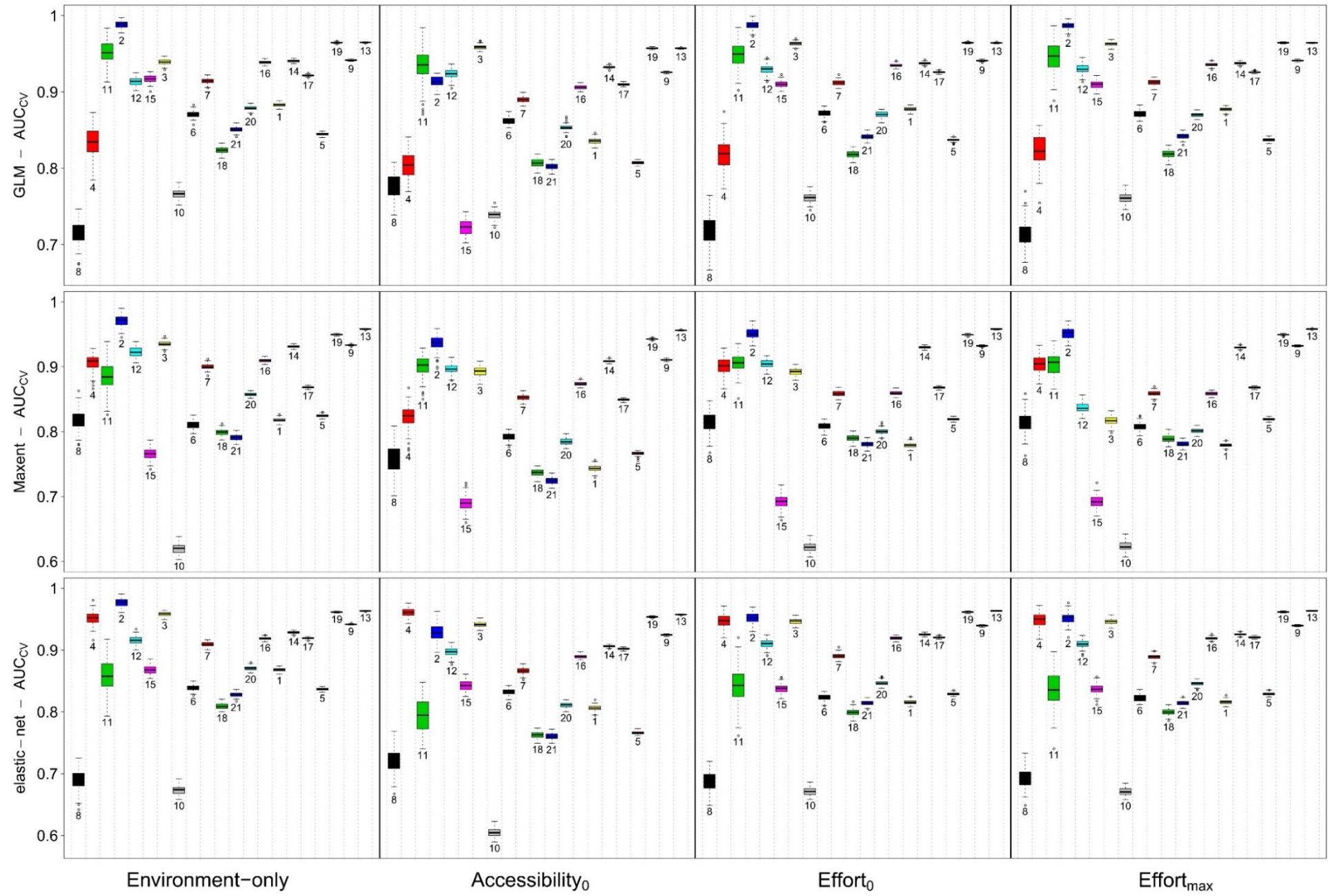
Panels a & c show results for the environment-only model (without any bias manipulation) and bias-accounting models (accessibility and effort models) without correcting for sampling bias. However, panels b & d compare results for the environment-only model and bias-accounting models (accessibility and effort models) after correcting for sampling bias (see main text; for effort models, plots show evaluations either conditioning the predictions on a value of zero or the maximum relative sampling effort of training presences). Species are in ascending order according to their number of occupied pixels at cross-validation scale (with numbers represent the species; see Table 1 for full species names).

**Fig. A10:** Similar to Fig. A9, but showing results of independent evaluation in Egypt. Evaluation metrics are calculated in Egypt using mean predictions (of 5-folds cross-validation) in Egypt along with entirely independent species records from Egypt (not used to run any of the models). Species are in ascending order according to their number of occupied pixels in Egypt (with numbers represent the species; see Table 1 for full species names).

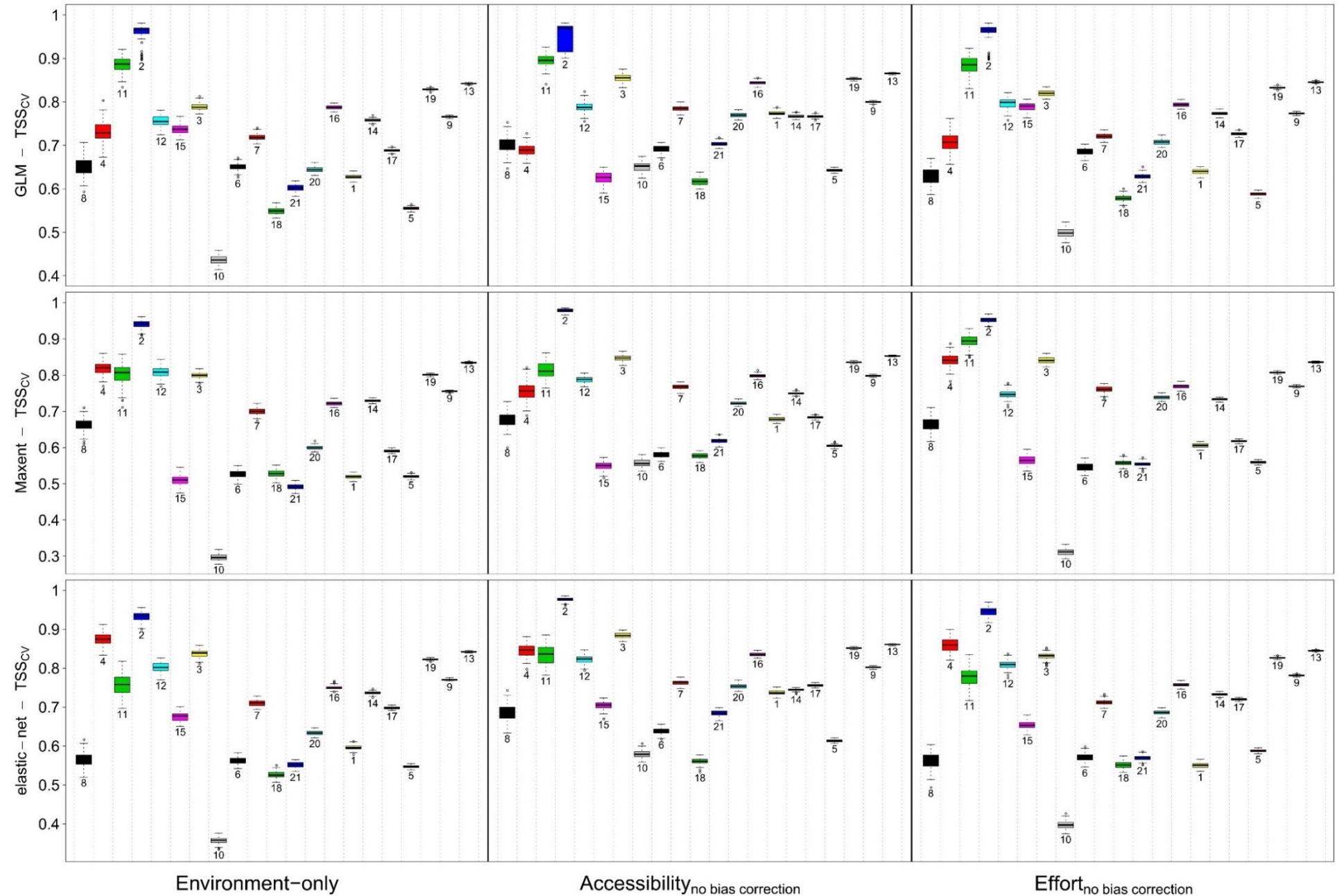
**Fig. A11:** Species mean TSS calculated either on cross-validation (a - b) or in Egypt (c - d), either without (modelling evaluation; a & c) or with (bias-free evaluation; b & d) sampling bias correction (for details, see Appendix 6). Each species is represented by different symbols (similar to those shown in Fig. A7; numbers represent species used, see Table 1). Red lines indicate the overall mean TSS at each modelling algorithm and bias models applied.



**Fig. A9 (a)**



**Fig. A9 (b)**



**Fig. A9 (c)**

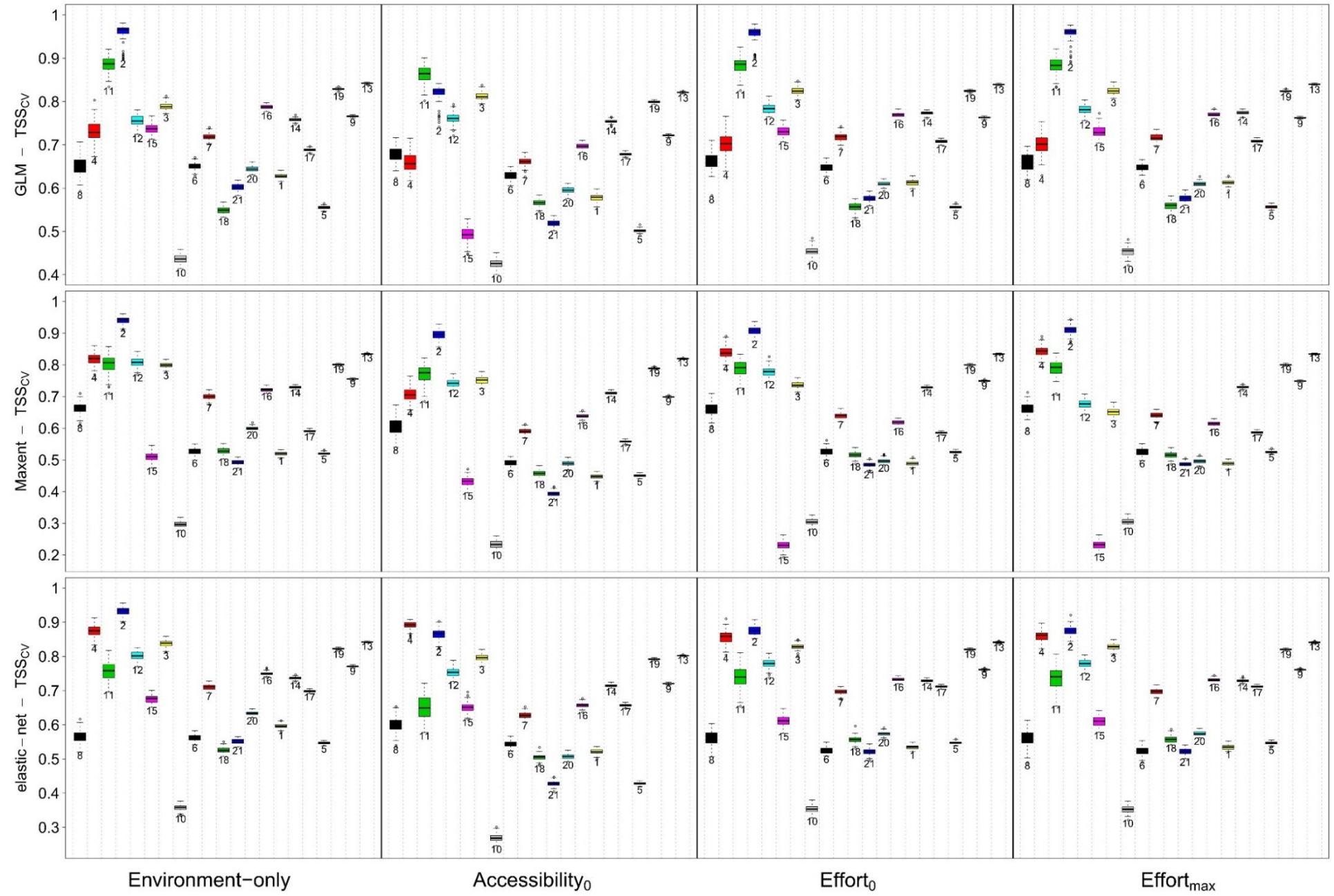
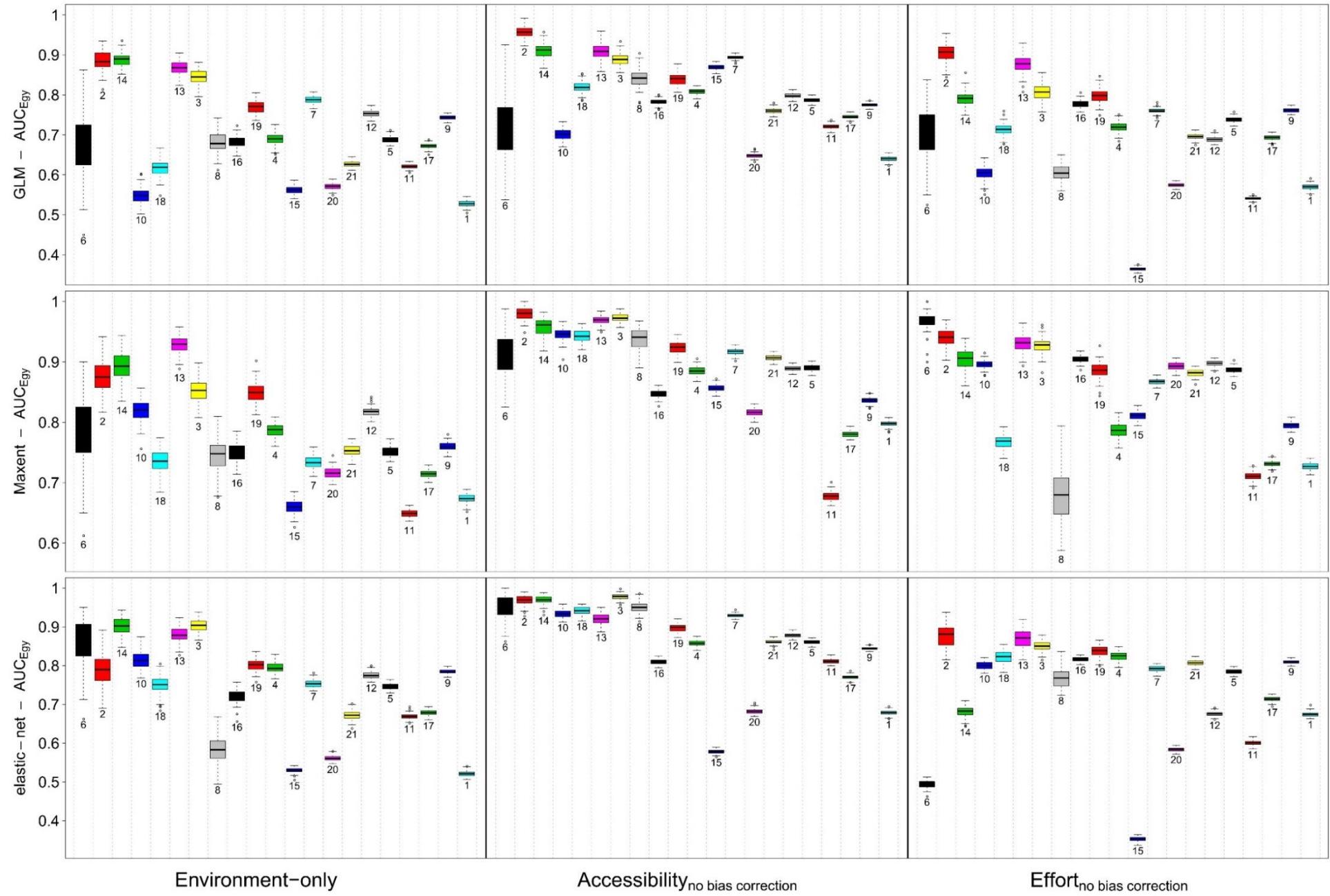
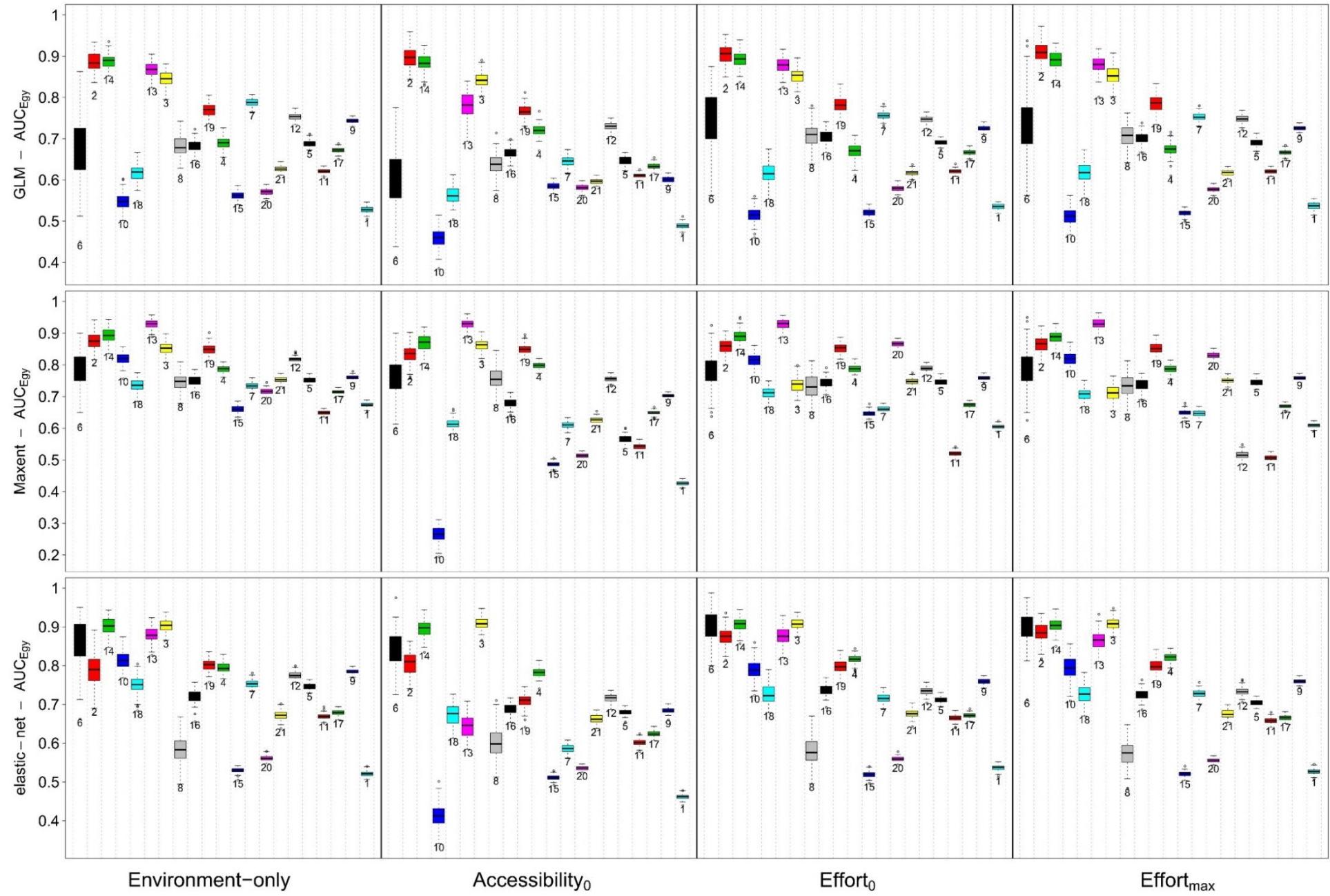


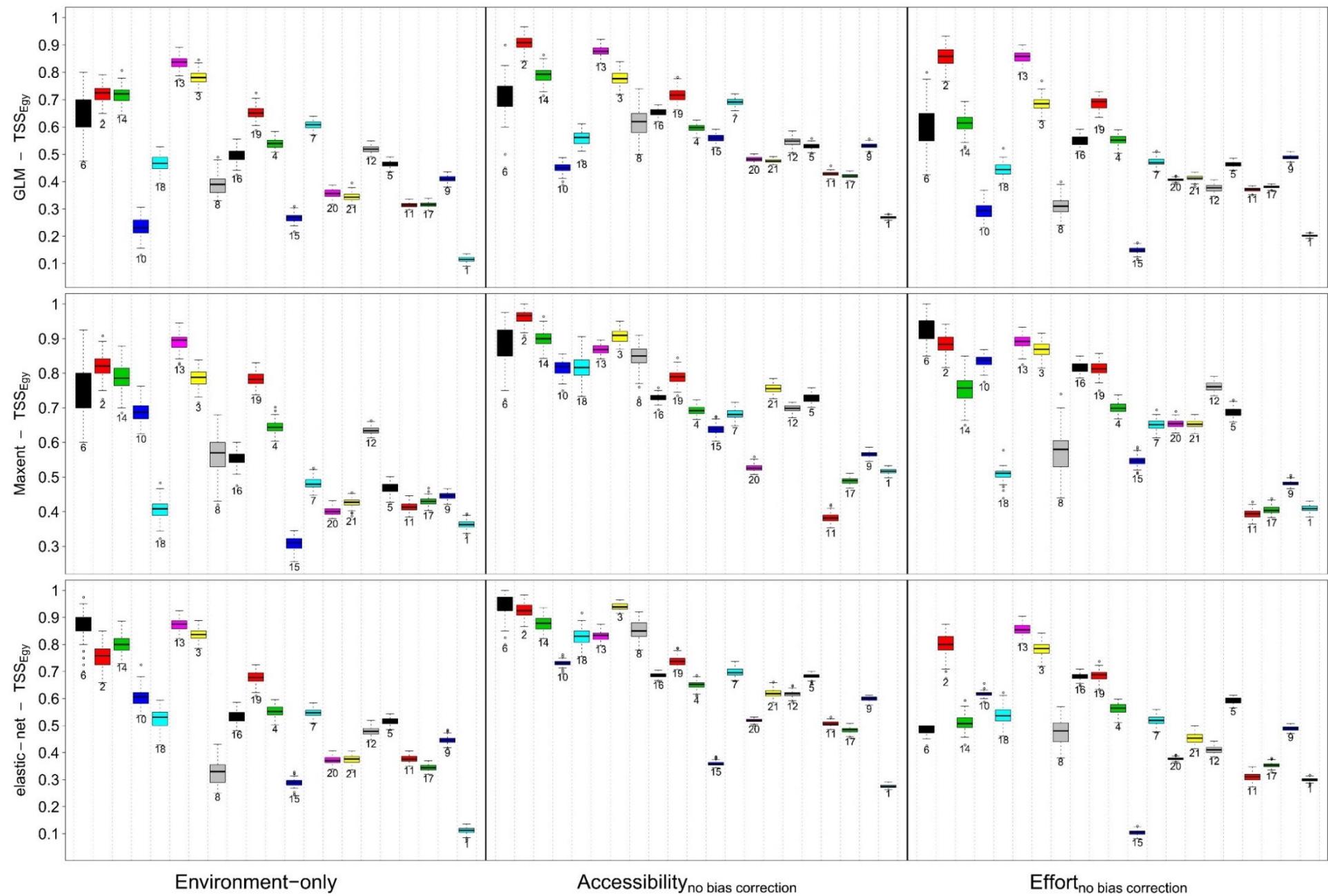
Fig. A9 (d)



**Fig. A10 (a)**



**Fig. A10 (b)**



**Fig. A10 (c)**

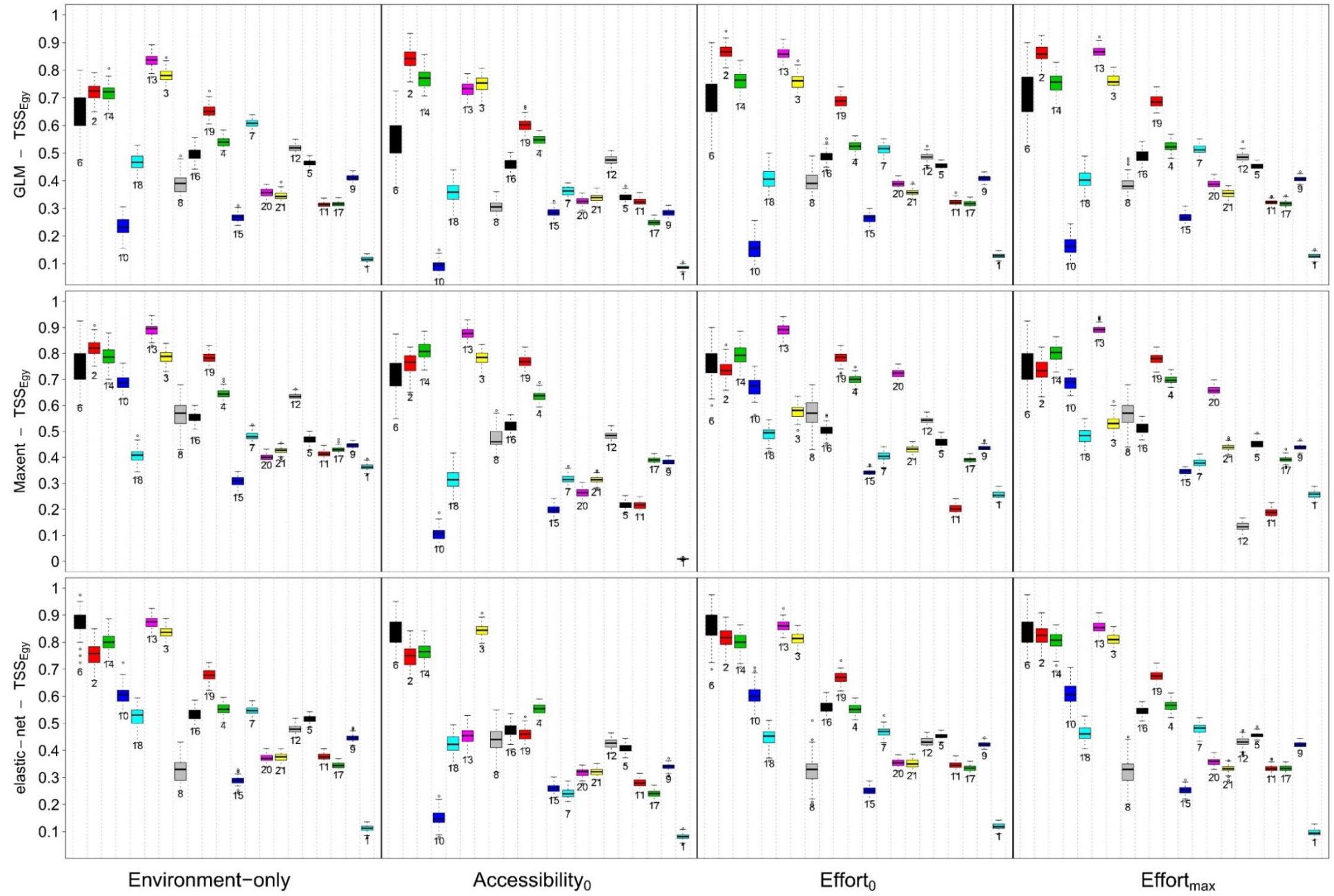
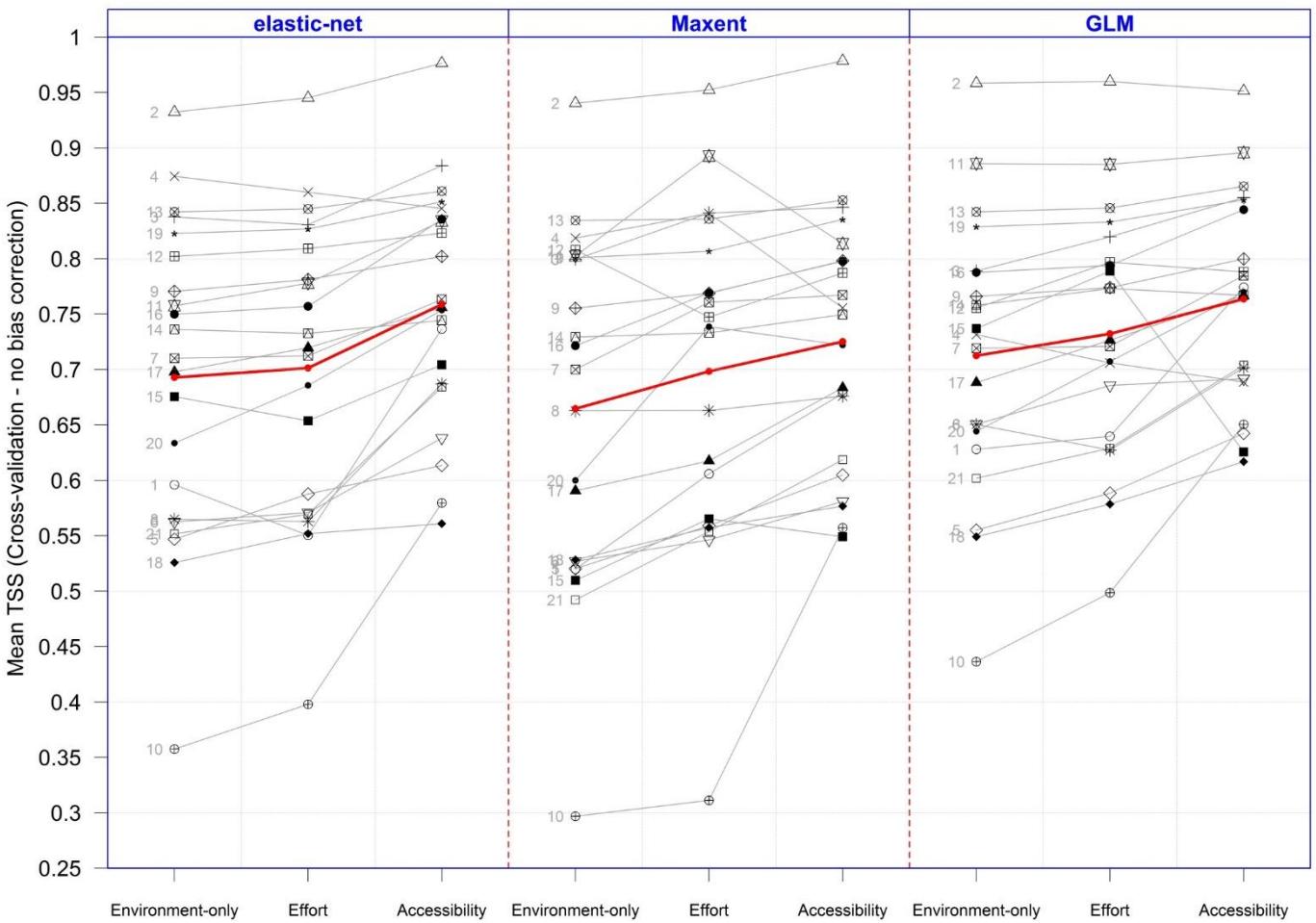
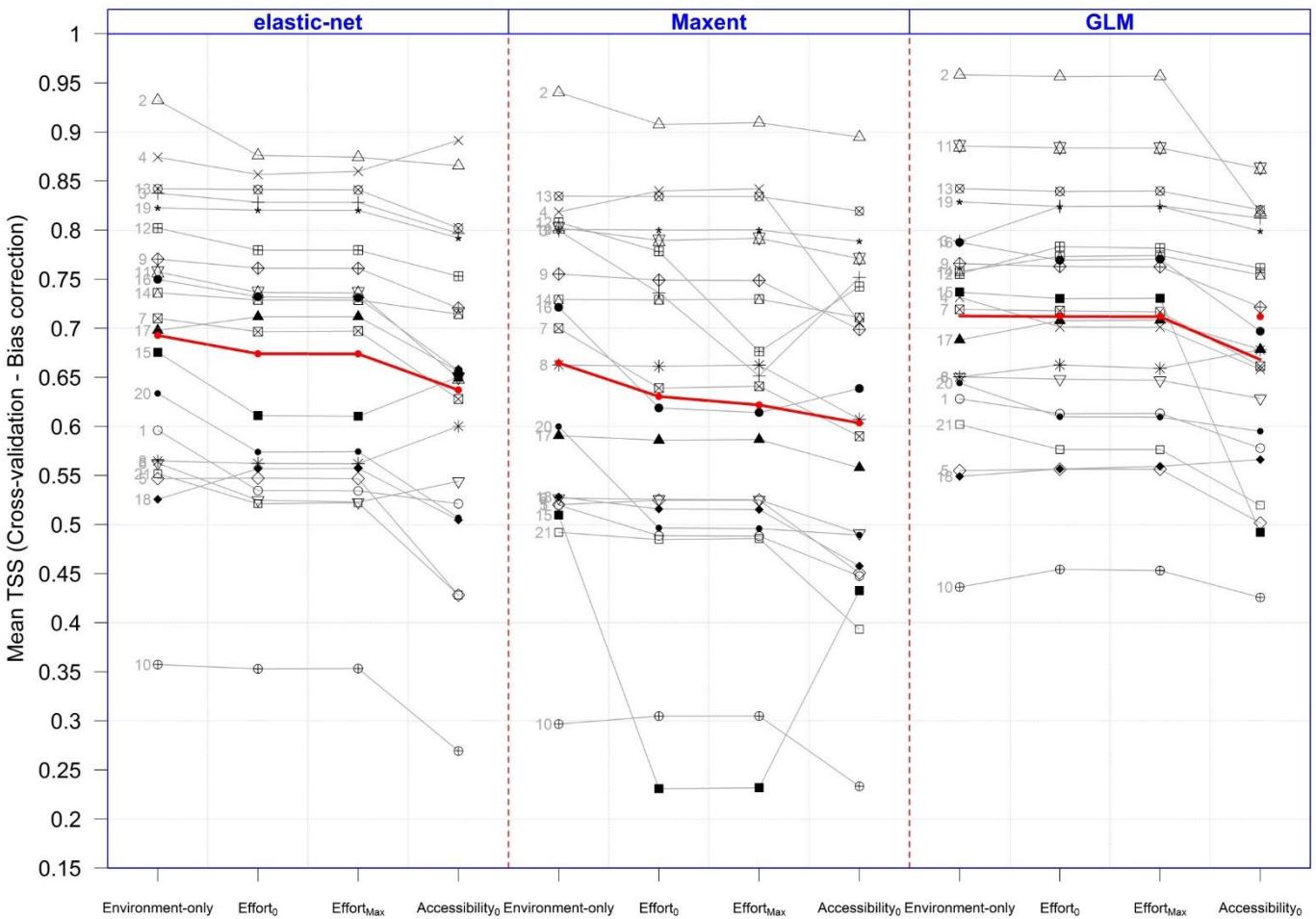


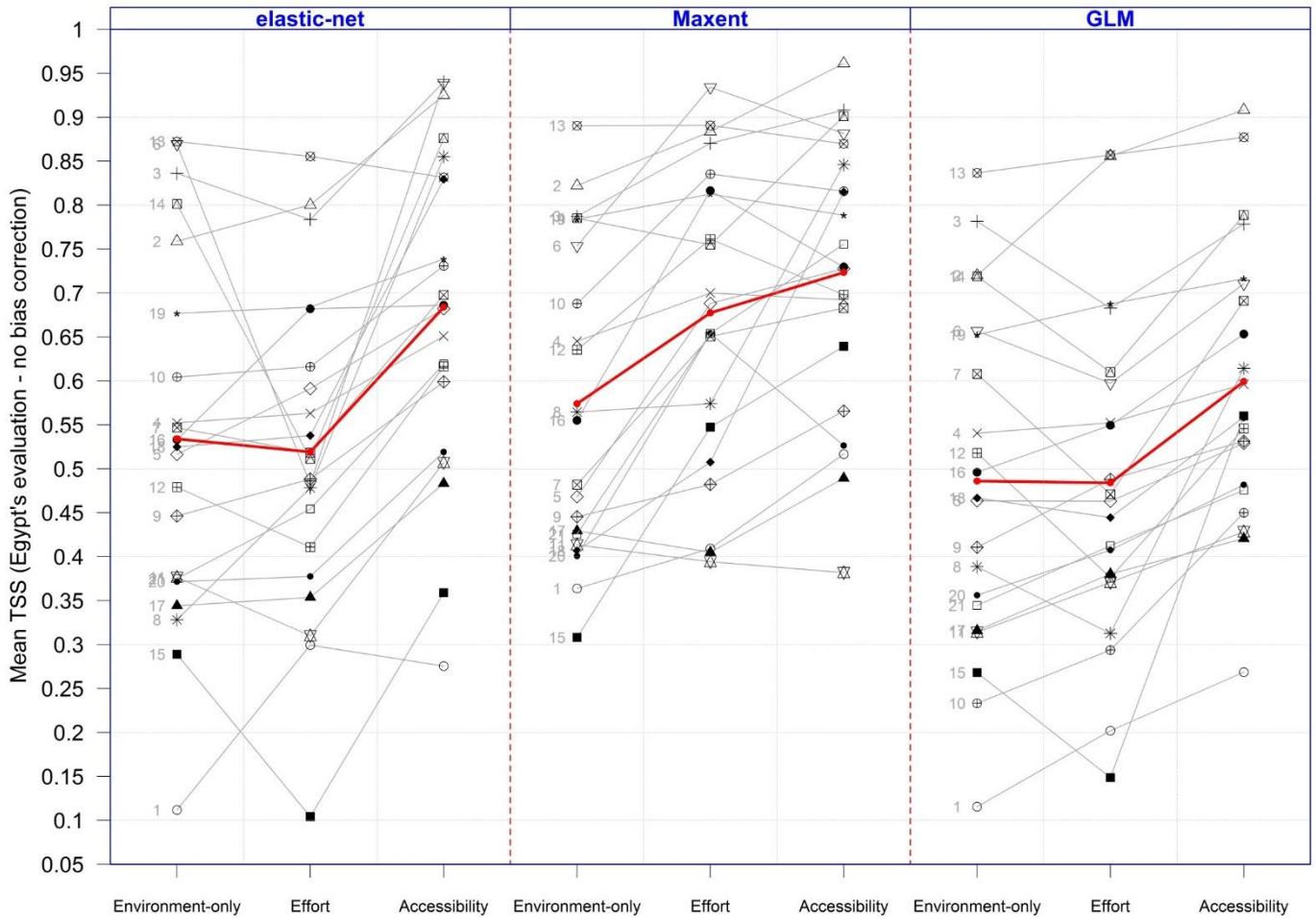
Fig. A10 (d)



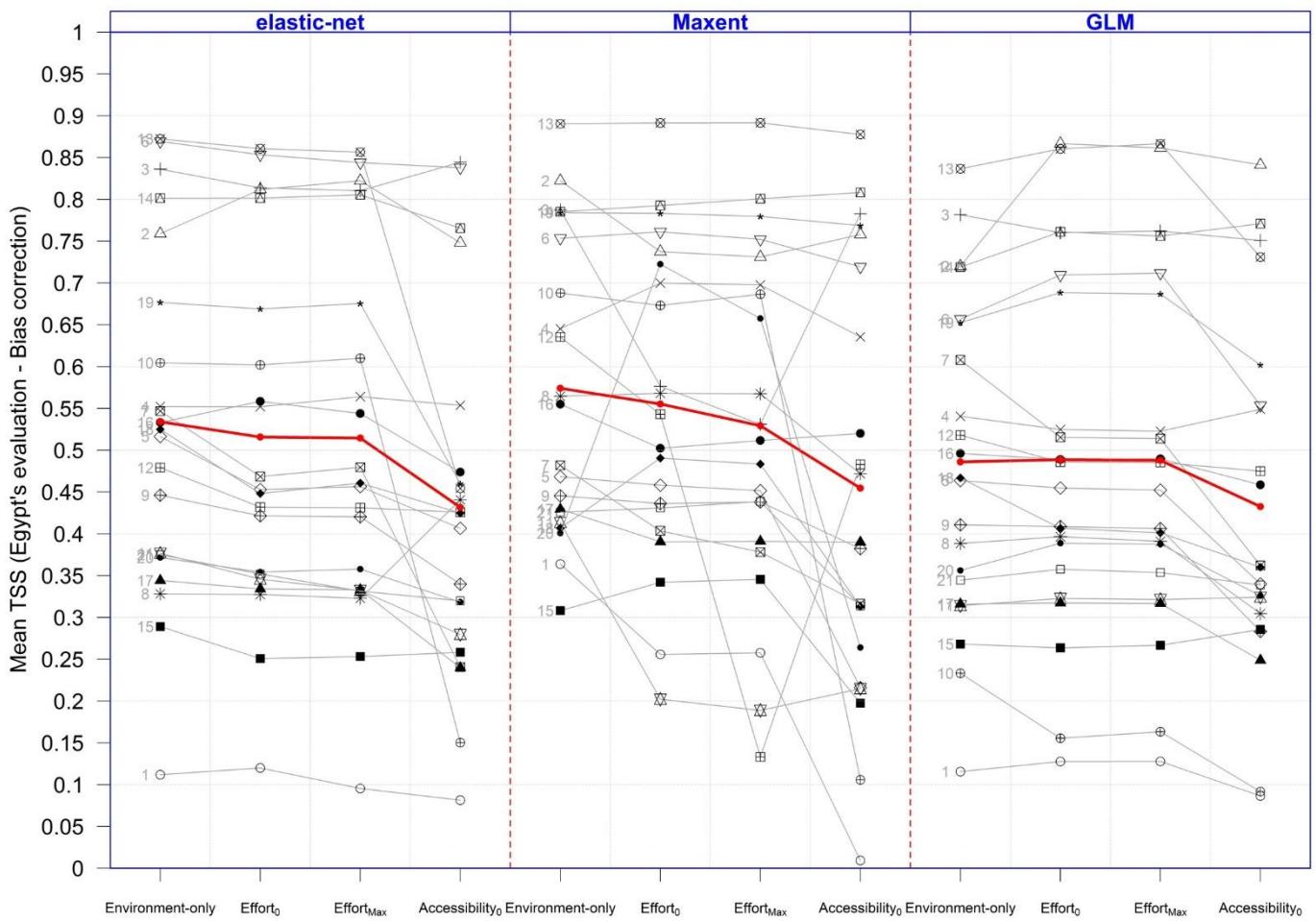
**Fig. A11 (a)**



**Fig. A11 (b)**



**Fig. A11 (c)**



**Fig. A11 (d)**

**Fig. A12:** Kendall's correlation of the per-species mean TSS between different pairs of modelling algorithms (same as Fig. 2 in the main text, but for TSS). Each species is represented by different symbol (similar to those of Fig. A7) with different colours for different bias models applied (using predictions of environment-only model and bias-free prediction of accessibility and effort model). 'M' indicates the overall mean evaluation. First row represents mean spatial-block cross-validation evaluation, while the second is for independent evaluations in Egypt.

**Fig. A13:** Values of bias variables at presences and available locations (backgrounds), at the per-species study area (A) or at local scale 'Egypt' (B). Rows correspond to different species, with numbers indicates the species (see Table 1), and columns represent bias variables of the accessibility model (distances to roads, cities, and protected areas) and effort model (relative bats' sampling intensity). For each species, values at presence locations are indicated with black, and values at pixels unoccupied by the species are shown in grey. Most of the species are recorded from closer to roads and cities (and to some extent, the protected areas), and unexpectedly at low to moderate sampling efforts.

**Fig. A14:** AUC scores calculated the default way (using all available testing backgrounds – y-axis) versus using a fixed ratio between testing presences and backgrounds (test-data prevalence = 1:20 – x-axis).

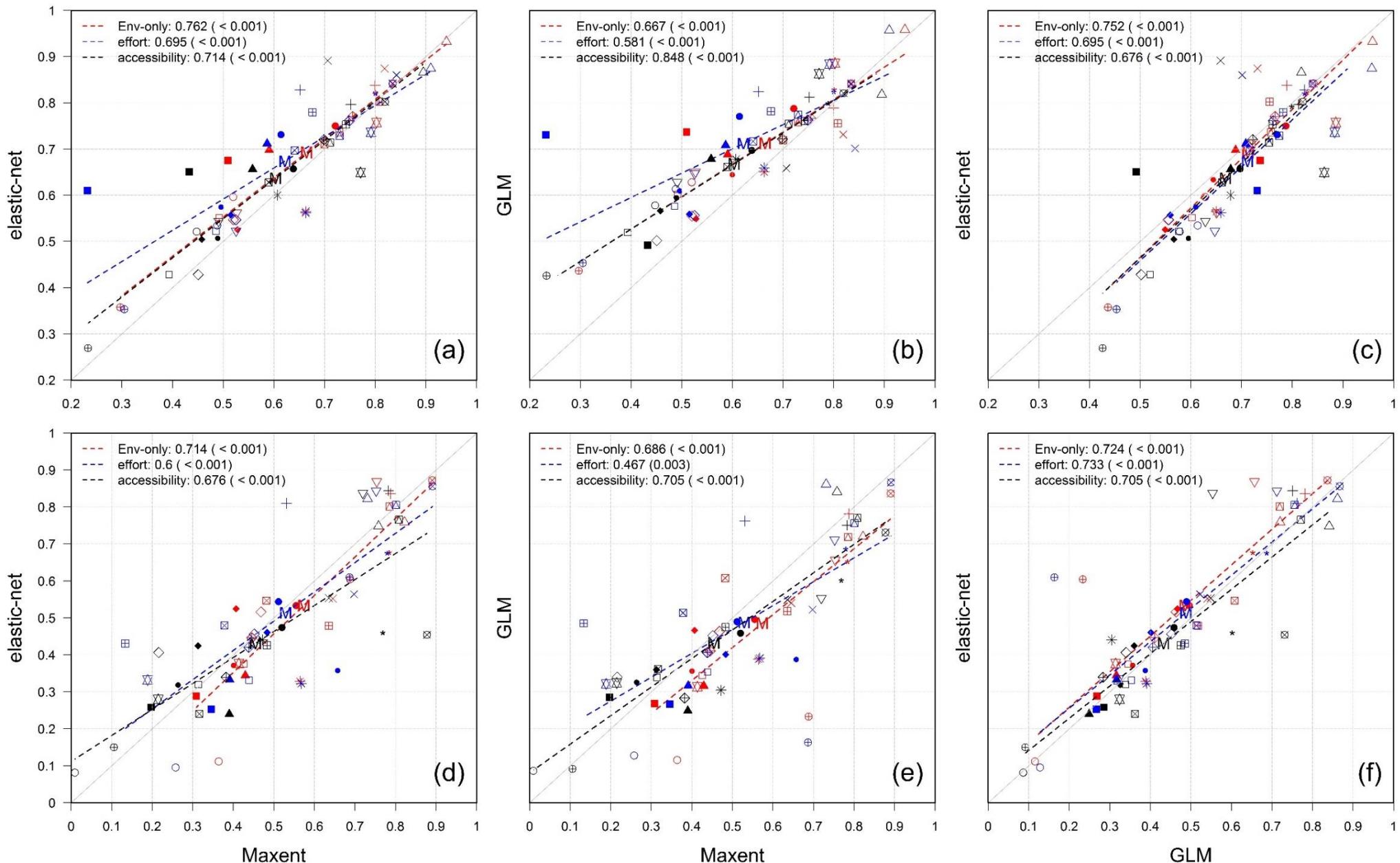
- (A) shows the raw outputs of 5-folds cross-validation.
- (B) shows the per-species mean AUC on cross-validation.

Different colours represent the three bias models applied (environment-only, accessibility, and efforts).

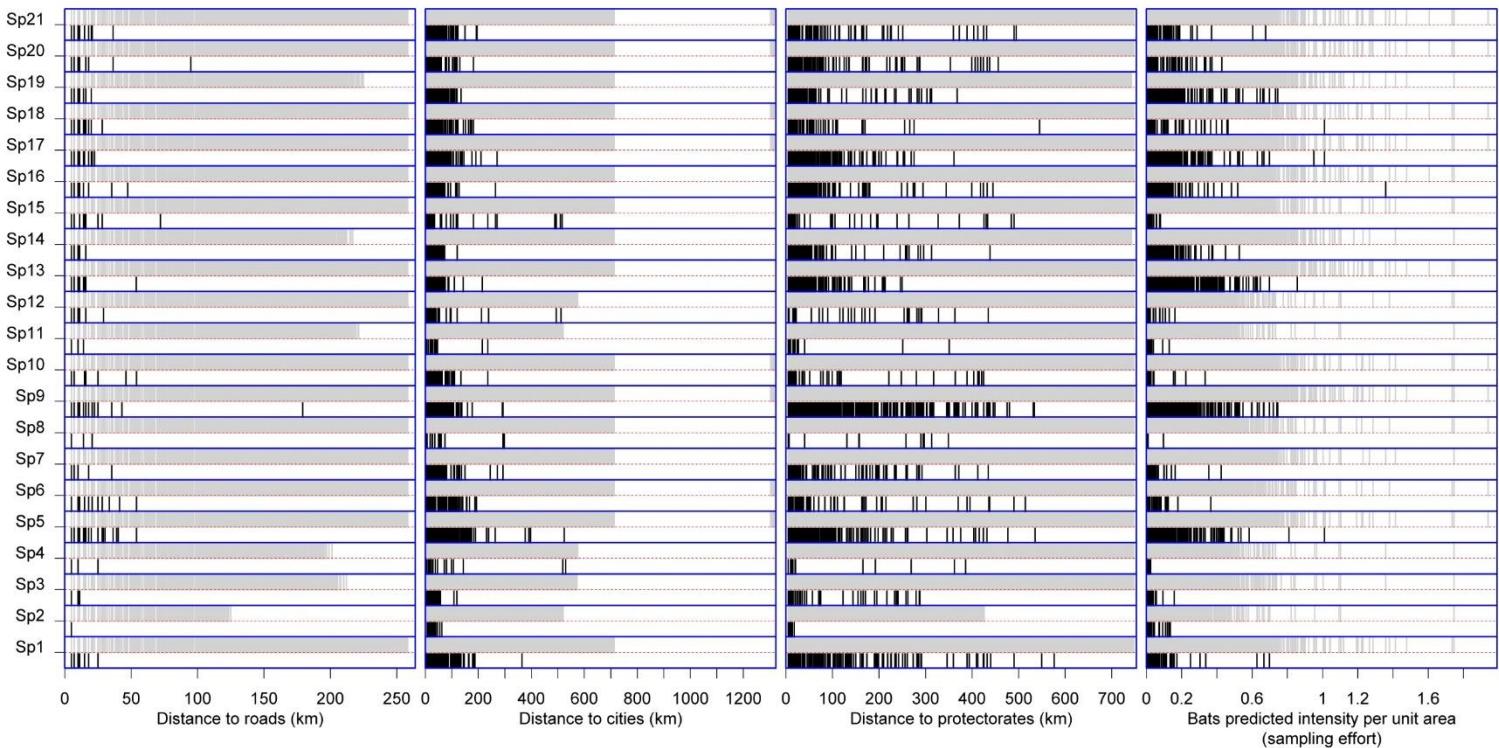
**Fig. A15:** The predicted distribution of *Otonycteris hemprichii* (mean of 5-folds cross-validation), using different modelling algorithms (rows) and bias models (columns). Maps were scaled between zero and one, as different modelling algorithms do not have the same scale, with blue colour indicates higher predicted relative intensity.

(A) shows cropped predicted distribution of *Otonycteris hemprichii* (the same as Fig. 5) to Egypt. Grey points (in the top left panel) represents available records used for independent evaluation presences in Egypt.

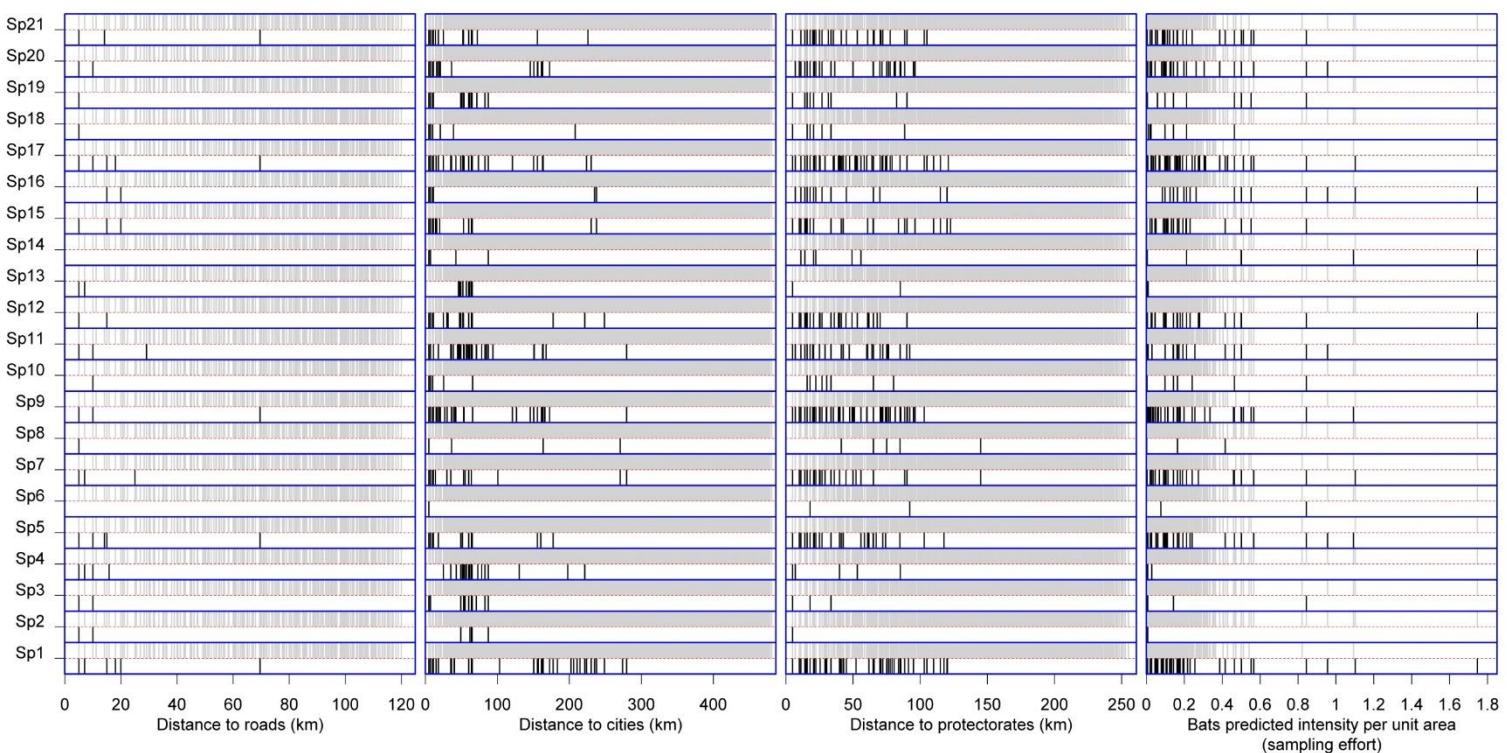
In Fig. 5 of the main text, extreme values ( $> 0.9995$  quantile of predicted values) were replaced with their next smaller value to improve map visualization, as GLM and elastic-net models sometimes yielded extreme values. Maps in (B) are equivalent to maps in Fig. 5, without any extreme values manipulation (using the linear scale), demonstrating the difficulty of visualizing the predicted patterns in the existence of extreme values; see main text for more details.



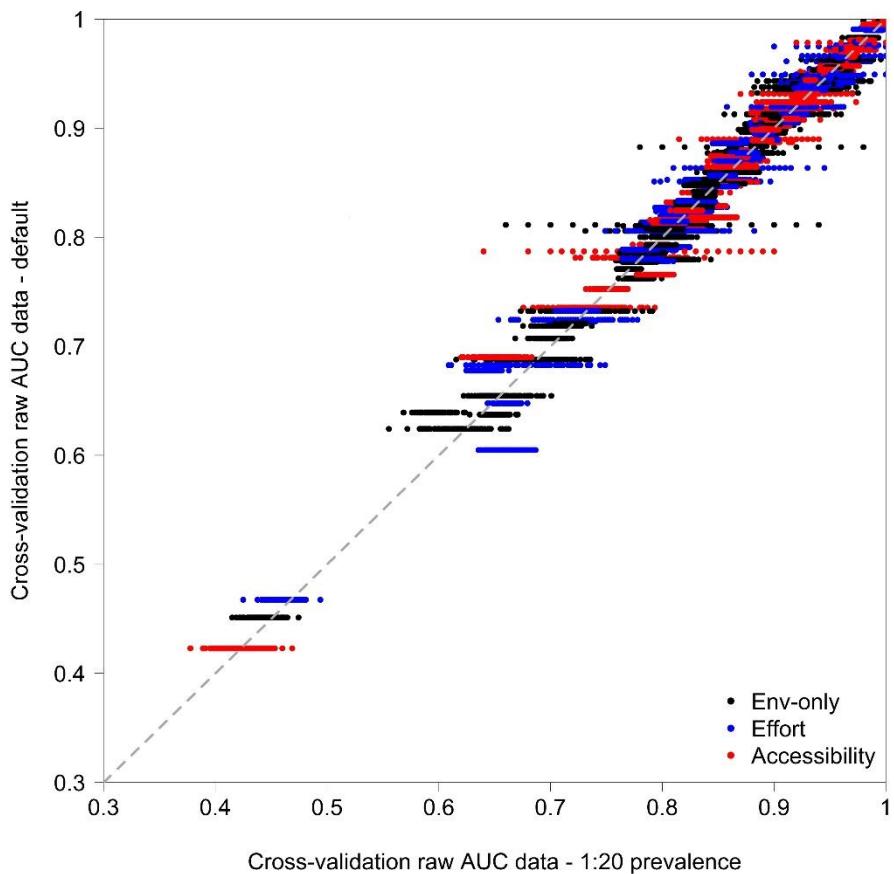
**Fig. A12**



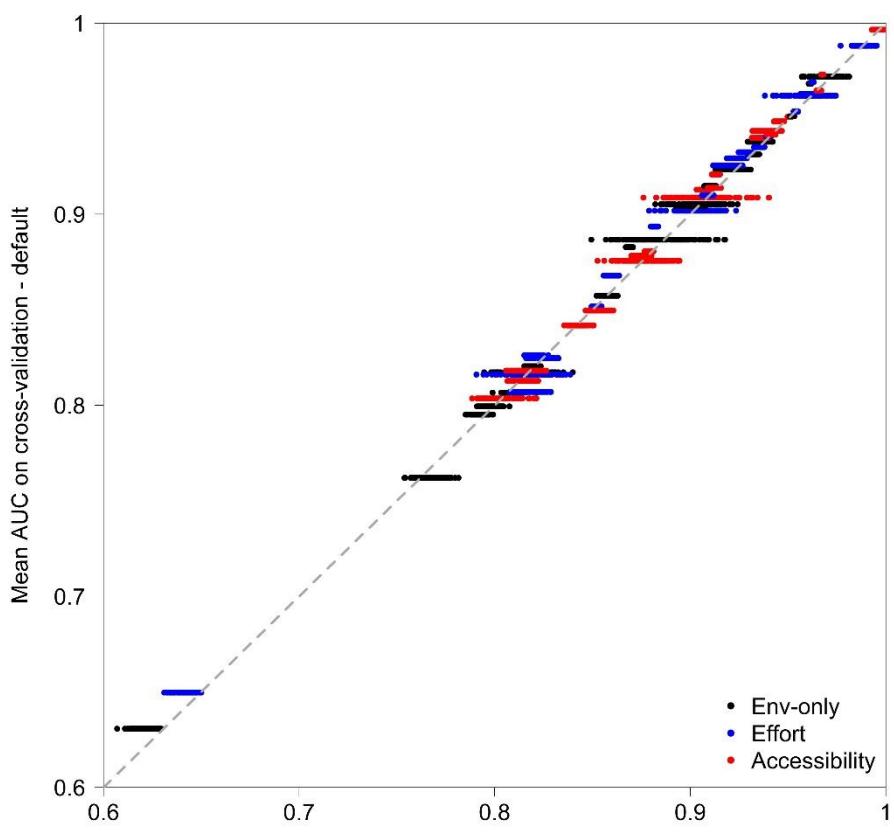
**Fig. A13 (A)**



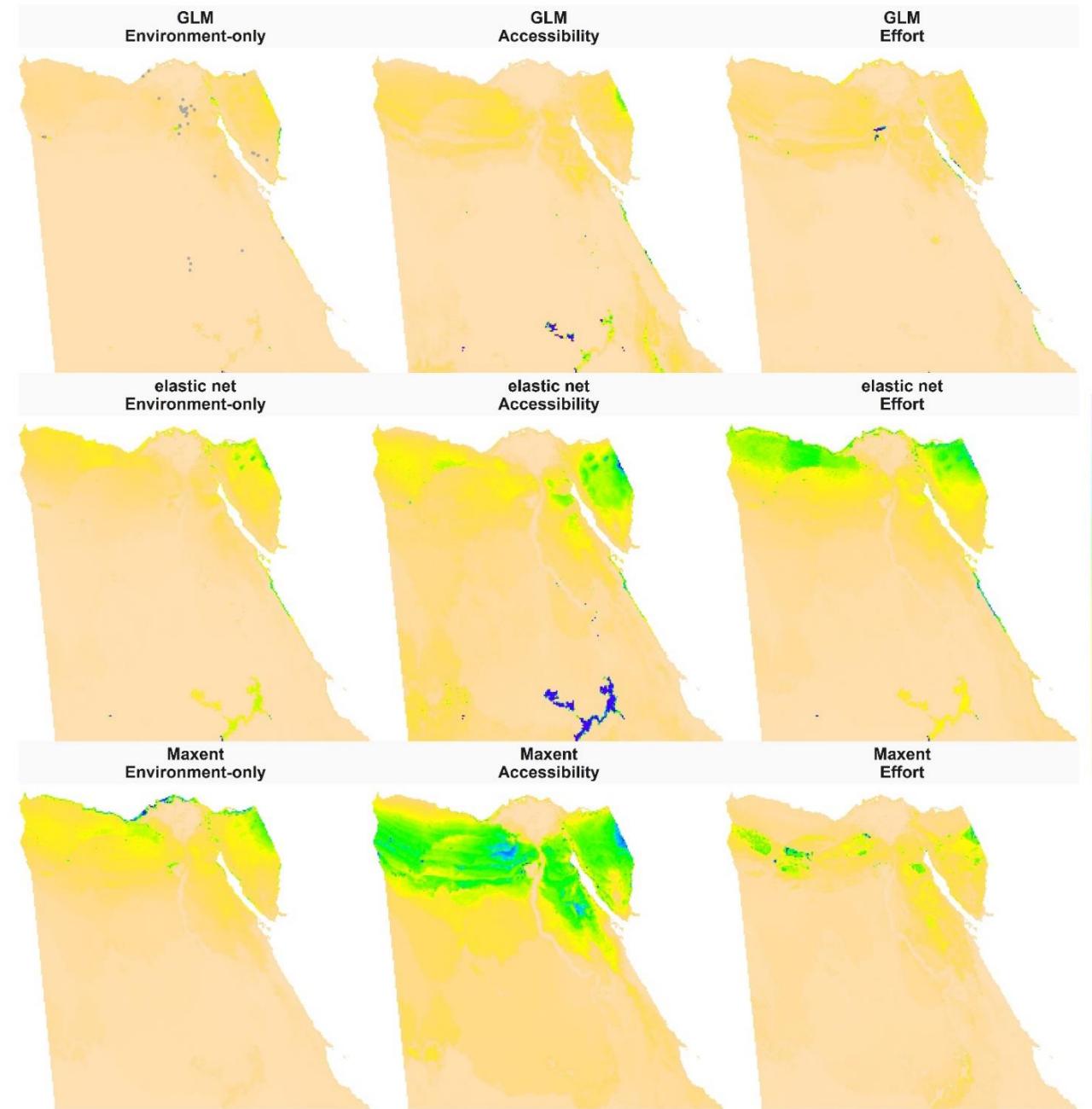
**Fig. A13 (B)**



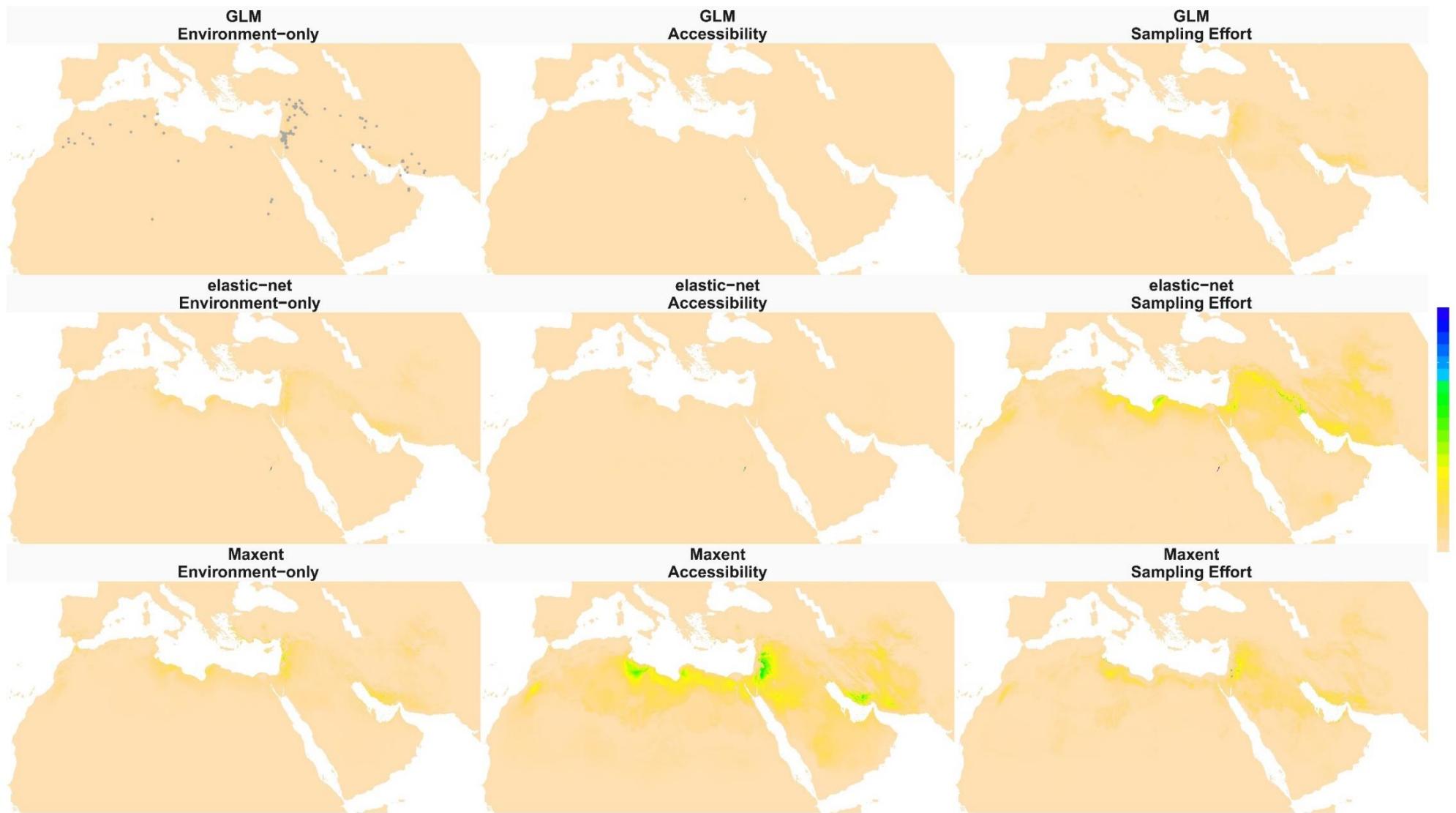
**Fig. A14(A)**



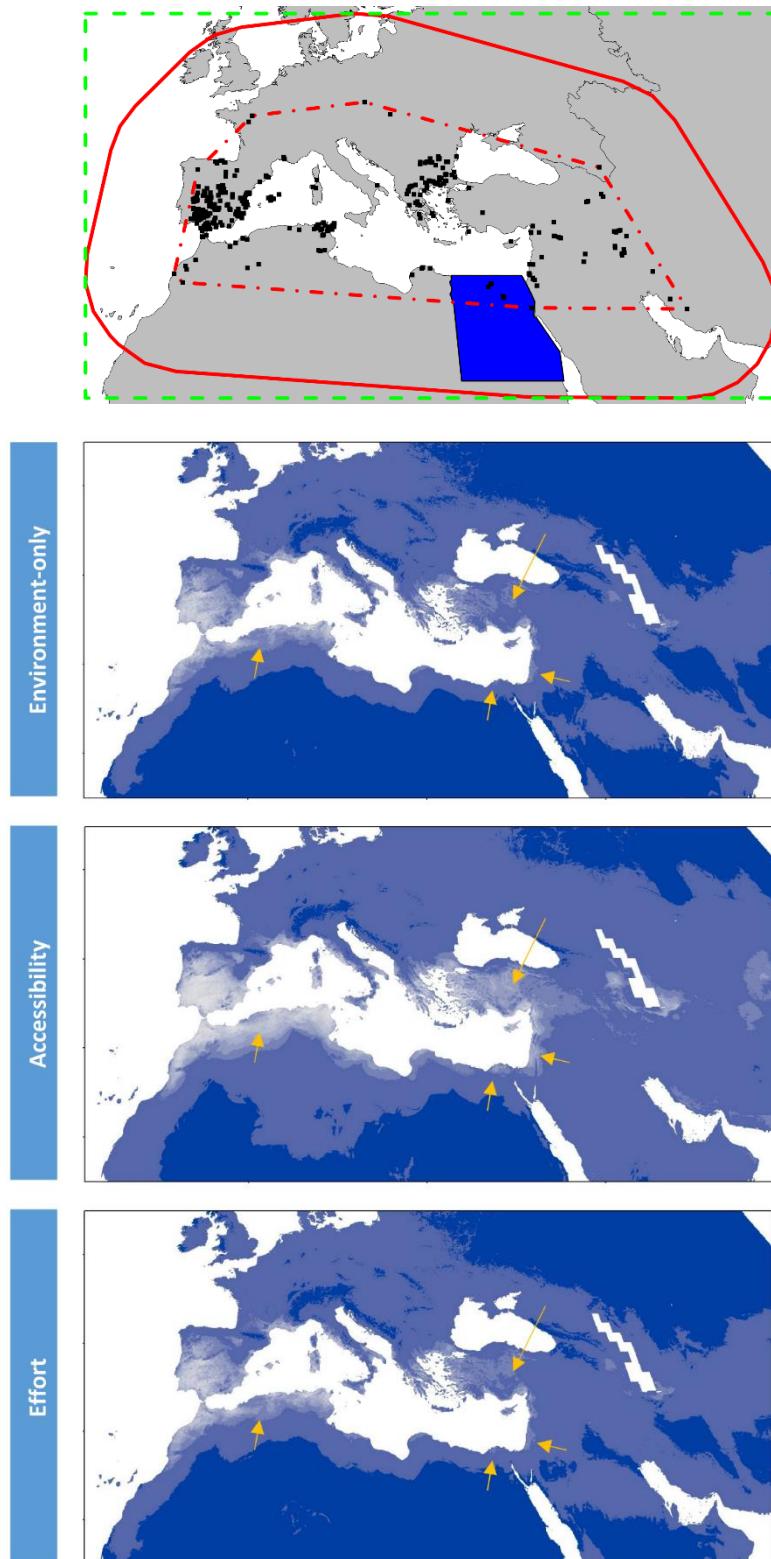
**Fig. A14(B)**



**Fig. A15 (A)**



**Fig. A15 (B)**



**Fig. A16:** The reported and predicted distribution of *Rhinolophus mehelyi* (Maxent). The 2<sup>nd</sup> to the 4<sup>th</sup> panels show predictions from environment-only model and bias-free prediction of accessibility and effort models, respectively. Arrows represent regions which gain higher predicted values after correcting for sampling bias (such as central Turkey or the Algerian Atlas). Predictions at these areas are of higher uncertainty (lower congruence) and field validation is probably required to confirm the species existence. The lighter the colour, the more suitable the location.

## Appendix 2: List of literature resources used for extracting bats records

- Abd Rabou A.-F.N., Yassin M.M., Al Agha M.R., Hamad D.M., Ali A.-K.S. (2007) **Wild mammals in the Gaza Strip, with particular reference to Wadi Gaza.** The Islamic University Journal (Series of Natural Studies and Engineering) 15(1):87-109.
- Albayrak I. (2003) **The bats of the eastern Black Sea region in Turkey (Mammalia: Chiroptera).** Turkish Journal of Zoology 27(4):269-273.
- Al-kuwari S.K. (1999) **On *Prosthodendrium parvouterus* (Trematoda: Lecithodendriidae) a parasite of the bat *Taphozous(sic) nudiventris*.** Qatar University Science Journal, 18:155-158.
- Allegrini B., Durand G., Durand E., Peyre O. (2011) **On some bats recorded in the Adrar region, Mauritania.** African Bat Conservation News, 26:2:4.
- Al-Omari K.A., Abu Baker M.A., Amr Z.S. (2000) **First record of the Egyptian Slitfaced Bat, *Nycteris thebaica*, from Jordan.** Zoology in the Middle East 21:5-7.
- Al-Shanti M.A., Pint J.J., Al-Juaid A.J., Al-Amoudi S.A. (2003) **Preliminary survey for caves in the Habakah region of the Kingdom of Saudi Arabia.** Saudi Geological Survey Open-File Report (SGS-OF-2003-3), 32 p.
- Anderson J. (1902) **Zoology of Egypt: Mammalia (compiled by W.E. de Winton).** London: Hugh Rees Ltd.
- Andrianivoarivelo R.A., Andriafidison D., Rahaingonirina C., Raharimbola S., Rakotoarivelo A.A., Ramilijaona O.R., Racey P.A., Jenkins R.K.B (2011) **A conservation assessment of *Rousettus madagascariensis* (Grandidier, 1929, Pteropodidae) roosts in eastern Madagascar.** Madagascar conservation & development 6(2):78-82.
- Angelici F.M., Wariboko S.M., Luiselli L., Politano E. (2000) **A long-term ecological survey of bats (Mammalia, Chiroptera) in the eastern Niger Delta (Nigeria).** Italian Journal of Zoology 67(2):169-174.
- Ansell W.F.H. (1986) **Some Chiroptera from south-central Africa.** Mammalia 50(4):507-520.
- Archer A.L. (1977) Results of the Winifred T. Carter expedition 1975 to Botswana: Mammals - Chiroptera. Botswana Notes & Records 9:145-154.
- Aulagnier S., Denys C. (2000) **Record of the naked-rumped bat *Taphozous nudiventris*, Chiroptera, Emballonuridae in Morocco.** Mammalia, 64: 116-118.
- Baker R.J., Davis B.L., Jordan R.G., Binous A. et al. (1974) **Karyotypic and morphometric studies of Tunisian mammals: bats.** Mammalia 38(4):695-710.
- Baydemir N.A., Albayrak I. (2006) **A Study on the Breeding Biology of Some Bat Species in Turkey (Mammalia: Chiroptera).** Turkish Journal of Zoology 30(1):103-110.
- Benda P., Abi-Said M., Bartonička T., Bilgin R., Faizolahi K., Lučan R.K., Nicolaou H., Reiter A., Shohdi W.M., Uhrin M., Horáček I. (2011) ***Rousettus aegyptiacus* (Pteropodidae) in the Palaearctic: list of records and revision of the distribution range.** Vespertilio, 15: 3-36.
- Benda P., Al-Juaid M.M., Reiter A., Nasser A.K. (2011) **Noteworthy records of bats from Yemen with description of a new species from Socotra.** Hystrix, n.s., 22(1):23-56.
- Benda P., Andreas M., Kock D., Lučan R., Munclinger P., Nová P., Obuch J., Ochman K., Reiter A., Uhrin M., Weinfurtová D. (2006) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean. Part 4. Bat fauna of Syria: distribution, systematics, ecology.** Acta Societatis Zoologica Bohemicae, 70:1-329.
- Benda P., Andriollo T., Ruedi M. (2014) **Systematic position and taxonomy of *Pipistrellus deserti* (Chiroptera: Vespertilionidae).** Mammalia.
- Benda P., Červený J., Konečný A., Reiter A., Ševčík M., Uhrin M., Vallo P. (2010) **Some new records of bats from Morocco (Chiroptera).** Lynx 41:151-166.
- Benda P., Dietz C., Andreas M., Hotový J., Lučan R., Maltby A., Meakin K., Truscott J., Vallo P. (2008) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean and Middle East. Part 6. Bats of Sinai (Egypt) with some taxonomic, ecological and echolocation data on that fauna.** Acta Societatis Zoologicae Bohemicae 72(1-2):1-103.

- Benda P., Faizolâhi K., Andreas M., Obuch J., Reiter A., Ševčík M., Uhrin M., Vallo P., Ashrafi S. (2012) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean and Middle East. Part 10. Bat fauna of Iran.** Acta Societatis Zoologicae Bohemicae, 76:163–582.
- Benda P., Georgiakakis P., Dietz Ch., Hanák V., Galanaki K., Markantonatou V., Chudárková A., Hulva P., Horáček I. (2012) **Bats (Mammalia: Chiroptera) of the Mediterranean and Middle East. Part 7. The bat fauna of Crete, Greece.** Acta Societatis Zoologicae Bohemicae 72:105-190.
- Benda P., Gvoždík V. (2010) **Taxonomy of the genus *Otonycteris* (Chiroptera: Vespertilionidae: Plecotini) as inferred from morphological and mtDNA data.** Acta Chiropterologica, 12(1):83-102.
- Benda P., Hanák V., Červený J. (2011) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean and Middle East. Part 9. Bats from Transcaucasia and West Turkestan in collection of the National Museum, Prague.** Acta Societatis Zoologicae Bohemicae, 75:159–222.
- Benda P., Hanák V., Andreas M., Reiter A., Uhrin M. (2004) **Two new species of bats (Chiroptera) for the fauna of Libya: *Rhinopoma hardwickii* and *Pipistrellus rueppellii*.** Myotis, 41–42: 109-124.
- Benda P., Hanák V., Horáček I., Hulva P., Lučan R., Ruedi M. (2007) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean. Part 5. Bat fauna of Cyprus: review of records with confirmation of six species new for the island and description of a new subspecies.** Acta Societatis Zoologicae Bohemicae, 71: 71–130.
- Benda P., Ivanova T., Horáček I., Hanák V., Červený J., Gaisler J., Gueorguieva A., Petrov B., Vohralík V. (2003) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean. Part 3. Review of bat distribution in Bulgaria.** Acta Societatis Zoologicae Bohemicae, 67:245–357.
- Benda P., Kiefer A., Hanák V., Veith M. (2004) **Systematic status of African populations of long-eared bats, genus *Plecotus* (Mammalia: Chiroptera).** Folia Zoologica, 53(1):1–48.
- Benda P., Lučan R. K., Shohdi W. M., Porteš M., Horáček I. (2014) **Microbats of the Western Oases of Egypt, Libyan Desert.** Vespertilio, 17:45–58.
- Benda P., Lučan R.K., Obuch J., Reiter A., Andreas M., Baćkor P., Bohnenstengel T., Eid E.K., Ševčík M., Vallo P., et al. (2010) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean and Middle East. Part 8. Bats of Jordan: fauna, ecology, echolocation, ectoparasites.** Acta Societas Zoologicae Bohemicae, 74:185–353.
- Benda P., Reiter A., Al-Jumaily M., Nasher A.-K., Hulva P. (2009) **A new species of mouse-tailed bat (Chiroptera: Rhinopomatidae: *Rhinopoma*) from Yemen.** Journal of the National Museum (Prague), Natural History Series 177 (6):53–68.
- Benda P., Ruedi M. (2004) **New data on the distribution of bats (Chiroptera) in Morocco.** Lynx, n. s., 35:13-44.
- Benda P., Spitzemberger F., Hanák V., Andreas M., Reiter A., Ševčík M., Šmid J., Uhrin M. (2014) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean and Middle East. Part 11. On the bat fauna of Libya II.** Acta Societatis Zoologicae Bohemicae, 78:1–162.
- Benda P., Vallo P. (2012) **New look on the geographical variation in *Rhinolophus clivosus* with description of a new horseshoe bat species from Cyrenaica, Libya.** Vespertilio, 16: 69–96.
- Benda P., Vallo P., Hulva P., Horáček I. (2012) **The Egyptian fruit bat *Rousettus aegyptiacus* (Chiroptera: Pteropodidae) in the Palaearctic: Geographical variation and taxonomic status.** Biologia, 67(6):1230–1244.
- Benda P., Vallo P., Reiter A. (2011) **Taxonomic revision of the genus *Asellia* (Chiroptera: Hipposideridae) with a description of a new species from Southern Arabia.** Acta Chiropterol 13(2):245–270.
- Bergmans W. (1979) **Taxonomy and zoogeography of the fruit bats of the People's Republic of Congo with notes on their reproductive biology (Mammalia, Megachiroptera).** Bijdragen tot de Dierkunde 48:161-186.
- Bergmans W., Bellier L., Vissault J. (1974) **A taxonomical report on a collection of Megachiroptera (Mammalia) from the Ivory Coast.** Revue Zoologique africaines 88:18-48
- Bilushenko A.A. (2013) **The Current Status of Kuhl's Pipistrelle, *Pipistrellus kuhlii* (Chiroptera, Vespertilionidae), in the Central Forest-Steppe of Ukraine.** Vestnik Zoologii, 47(4):343–349.
- Bonhote J.L. (1909) **On a Small Collection of Mammals from Egypt.** Proceedings of The Zoological Society of London, 79(4)788-802.

- Bontadinal F., Arlettaz R., Fankhauser T., Lutz M., Muhlethaler E., Theiler A., Zingg P. (2000) **The lesser horseshoe bat *Rhinolophus hipposideros* in Switzerland: present status and research recommendations.** Swiss Coordination Center for the Study and Protection of Bats, Zürich and Geneva.
- Cel'uch M., Ševčík M. (2006) **First record of *Pipistrellus kuhlii* (Chiroptera) from Slovakia.** Biologia 61(5):637-638.
- Churcher C.S. (1991) **The Egyptian fruit bat *Rousettus aegyptiacus* in Dakhleh Oasis, Western Desert of Egypt.** Mammalia, 551:139-143.
- Cockle A., Kock D., Stublefield L., Howell K.M., Burgess N.D. (1998) **Bat assemblages in Tanzanian coastal forests.** Mammalia 62(1):53-68.
- Cowan P. (2013) **An Annotated Checklist of the Mammals of Kuwait.** Sultan Qaboos University Journal for Science 18:19-24.
- D'Cruze N.C., McDonell Z.J., Langeveld T., Brooks P.D., Combrink X., Seemark E.C.J., Kearney T.C. (2008) **Bat survey in the uMkhuze section of the Greater St Lucia Wetland Park, KwaZulu-Natal, South Africa.** African Bat Conservation News 17:49.
- Dalhoumi R., Hedfi A., Aissa P., Aulagnier, S. (2014) **Bats of Jebel Mghilla National Park (central Tunisia): first survey and habitat-related activity.** Tropical Zoology 27(2):53-62.
- Darweesh N., Al-Melhim W.N., Disi A.M., Amr Z.S. (1997) **First record of the Naked-bellied Tomb Bat, *Taphozous nudiventris* Cretzschmar, 1830, from Jordan.** Zoology in the Middle East 15(1):13:14.
- Davy C.M., Russo D., Fenton M.B. (2007) **Use of native woodlands and traditional olive groves by foraging bats on a Mediterranean island: consequences for conservation.** Journal of Zoology 273(4):397-405.
- DeBlase A.F. (1971) **New distributional records of bats from Iran.** Fieldiana Zoology 58(3):9-14
- Decher J., Norris R.W., Fahr J. (2010) **Small mammal survey in the upper Seli River valley, Sierra Leone.** Mammalia 74(2):163–176.
- Dietz C. (2005): **Bats Final Report - Operation Wallacea.** Sinai 2005. 23 pp.
- Dornburg A., Colosi J.G., Maser C., Reese1 A.T., Watkins-Colwell G.J. (2011) **A survey of the Yale Peabody museum collection of Egyptian mammals collected during construction of the Aswan high dam, with an emphasis on material from the 1962–1965 Yale University prehistoric expedition to Nubia.** Bulletin of the Peabody Museum of Natural History 52(2):255-272.
- Dragu A., Munteanu I., Olteanu V. (2007) **First record of *Pipistrellus kuhlii* Kuhl, 1817 (Chiroptera: vespertilionidae) from Dobrogea (Romania).** Archives of Biological Sciences, 59(3):243-247.
- Eltringham S.K., Morley R.J., Kingdon J., Coe, M.J., MacWilliam N.C. (1999) **Checklist: mammals of Mkomazi.** In Coe, M.J. et al. 1999 Mkomazi: the ecology, biodiversity and conservation of a Tanzanian savanna. London, Royal Geographical Society, pp. 1-750.
- Etemad E. (1967) **Notes on some bats from Iran.** Mammalia 31(2):275–280.
- Fahr J., Djossa B.A., Vierhaus H. (2006) **Rapid assessment of bats (Chiroptera) in Déré, Diécké and Mt. Béro classified forests, southeastern Guinea; including a review of the distribution of bats in Guinée Forestière.** In H. E. Wright, J. McCullough, L. E. Alonso, M. S. Diallo (Eds.), A Rapid Biological Assessment of Three Classified Forests in Southeastern Guinea (pp. 168-180). Washington, D.C.: Conservation International.
- Fils E.M.B., Anonga A.G.B.A., Tsala D.B., Guiékéa B.B., Tsala D.B., Fotso A.K. (2014) **Diversity of bats of the far North Region of Cameroon – with two first records for the country.** Biodiversity 15(1):16-22.
- Flower S. S. (1932) **Notes on the recent mammals of Egypt, with a list of the species recorded from that kingdom.** Proceedings of the Zoological Society of London 1932: 369-450.
- Foui E. (2011) **Operation Wallacea: Sinai bats report.** Available at: [https://opwall.com/wp-content/uploads/OpWall\\_Sinai-bat-report\\_2011.pdf](https://opwall.com/wp-content/uploads/OpWall_Sinai-bat-report_2011.pdf)
- Freeman P.W. (1981) **A multivariate study of the family Molossidae (Mammalia, Chiroptera): morphology, ecology, evolution.** Fieldiana Zoology, New Series No. 7, Field Museum of Natural History, Chicago, 173 pp.
- Furman A., Özgül A. (2002) **Distribution of cave-dwelling bats and conservation status of underground habitats in the Istanbul area.** Ecological Research 17(1):69–77.

- Gaisler J., Madkour G., Pelikan J. (1972) **On the bats (Chiroptera) of Egypt.** Acta Scientiarum Naturalium Academiae Scientiarum Bohemicae, Brno 6(8):1-40.
- Gharaibeh B.M. (1997) **Systematics, distribution, and zoogeography of mammals of Tunisia.** Ph.D. thesis, Texas Tech University.
- Gharkheloo M.M., Karataş A., Kanlilic T. (2008) **Karyotype of *Pipistrellus kuhlii* (Natterer in Kuhl, 1819), (Chiroptera, Vespertilionidae) from Iran.** International Journal of Natural & Engineering Sciences 2(2):75.
- Ghassemi F., Kargar H., Nemati A. (2012) **The study of bat fauna in the south part of Iran: a case study of Jahrom.** Advances in Environmental Biology, 6(10):2720-2725.
- Godlevsky L., Tyshchenko V., Negoda V. (2000) **First Records of *Pipistrellus kuhlii* from Kyiv.** Vestnik Zoologii 34:78.
- Goiti U., Vecin P., Garin I., Salona M., Aihartza J.R. (2003) **Diet and prey selection in Kuhl's pipistrelle *Pipistrellus kuhlii* (Chiroptera: Vespertilionidae) in south-western Europe.** Acta Theriol. 48:457–468.
- Gomez G.O. (2012) **Establishing an isotopic basemap for flying foxes in Central Africa to assess the movement ecology of the straw-coloured fruit bat *Eidolon helvum* (Kerr, 1792).** Dissertation of the 2nd year, Magister in Ecology, Natural Heritage and Society (MNHN Paris)
- Hanák V., Benda P., Ruedi M. Horáček I., Sofianidou T.S. (2001) **Bats (Mammalia: Chiroptera) of the Eastern Mediterranean. Part 2. New records and review of distribution of bats in Greece.** Acta Societatis Zoologicae Bohemicae 65:279-346.
- Happold D.C.D., Happold M., Hill J.E. (1987) **The bats of Malawi.** Mammalia, 51:337–414.
- Harmsen R., Jabbal I. (1968) **Distribution and host-specificity of a number of fleas collected in south and central Kenya.** Journal of The East Africa Natural History Society and National Museum, XXVII(2):157-162.
- Harrison D.L. (1958) **A new race of tomb bat *Taphozous perforatus* E. Geoffroy, 1818, from northern Nigeria, with some observations on its breeding biology.** Durban Museum Novitates, 5:143-149.
- Harrison D.L. (1960) **A new species of Pipistrelle bat (Chiroptera: Pipistrellus) from south Israel.** Durban Museum Novitates 5(19):261-265.
- Harrison D.L. (1962) **A new bat for Israel, *Eptesicus innesi* Lataste, 1887, with some remarks on the affinities of this species.** Mammalian Biology (früher Zeitschrift für Säugetierkunde) 28:107-110.
- Harrison D.L. (1968) **On three Mammals new to the fauna of Oman, Arabia, with the description of a new subspecies of bat.** Mammalia 32(3):317-325.
- Harrison D.L. (1976) **Scientific results of the Oman flora and fauna survey, 1975. Description of a new subspecies of Botta's serotine (*Eptesicus bottae* Peters, 1869, Chiroptera: Vespertilionidae) from Oman.** Mammalia 39(3):415–418.
- Harrison D.L., Bates P.J.J. (1989) **Observations on two mammal species new to the Sultanate of Oman, *Vulpes cana* Blanford, 1877 (Carnivora: Canidae) and *Nycteris thebaica* Geoffroy, 1818 (Chiroptera: Nycteridae).** Bonner Zoologische Beiträge 40:73-77.
- Harrison D.L., Bates P.J.J. (1991) **The mammals of Arabia.** 2nd edition. Harrison Zoological Museum Publication.
- Harrison D.L., Makin D. (1988) **Significant new records of Vespertilionid Bats (Chiroptera : Vespertilionidae) from Israel.** Mammalia, 52(4):593–596.
- Harshey D.K., Chandra K. (2001) **Mammals of Madhya Pradesh and Chhattisgar.** Zoo's Print Journal, 16(2):659-668.
- Herzig-Straschil B., Robinson G.A. (1978) **On the ecology of the fruit bat, *Rousettus aegyptiacus* leachi (A. Smith, 1829) in the Tsitsikama Coastal National Park.** Koedoe, 21(1):101-110.
- Hoffmann M., Grubb P., Groves C., et al. (2009) **A synthesis of African and western Indian Ocean Island mammal taxa (Class: Mammalia) described between 1988 and 2008: An update to Allen (1939) and Ansell (1989).** Zootaxa, 2205:1-36.
- Horáček L., Benda P., Sadek R., Karkabi S., Abi-Said M., Lučan R., Hulva P., Karanouh R. (2008) **Bats of Lebanon, state of knowledge and perspectives.** Al-Ouat'Ouate, Revue Libanaise de Spéléologie et de Karstologie (N.S.), 14:52–67.

- Ifrim I., Valenciu N. (2006) ***Pipistrellus kuhlii* Kuhl, 1819, a new reported species for the chiropteran fauna of Moldavia (Romania).** Travaux du Muséum National d'Histoire Naturelle 'Grigore Antipa' 49:359–363.
- Jones C. (1971) **The Bats of Rio Muni, West Africa.** Journal of Mammalogy, 52(1):121-140.
- Juste J., Benda P., Garcia-Mudarra J.L., Ibáñez C. (2013) **Phylogeny and systematics of Old World serotine bats (genus *Eptesicus*, Vespertilionidae, Chiroptera): an integrative approach.** Zoologica Scripta, 42(5):441–457.
- Juste J., Ibáñez C. (1994) **Bats of the Gulf of Guinea islands: faunal composition and origins.** Biodiversity & Conservation 3(9):837-850.
- Karatas A., Sözen M. (2002) **Karyotype of *Taphozous nudiventris* Cretzschmar, 1830 (Mammalia: Chiroptera) from Turkey.** Israel Journal of Zoology, 48(4):359-360.
- Karatas A., Sözen M. (2006) **Bats of the middle and upper Kizilirmak regions, Central Anatolia, Turkey.** Lynx (N.S.), 37:1–268.
- Kityo R., Kerbis J.C. (1996) **Observations on the distribution and ecology of bats in Uganda.** Journal of East African Natural History, 85(1):49-63.
- Kock D., Barnett L., Fahr J., Emms C. (2002) **On a collection of bats (Mammalia: Chiroptera) from the Gambia.** Acta Chiropterologica, 4(1):77-97.
- Koopman K.F. (1975) **Bats of the Sudan.** Bulletin of the American Museum of Natural History 154(4).
- Koopman K.F. (1989) **Systematic Notes on Liberian Bats.** American Museum novitates, no. 2946.
- Koopman K.F., Mumford R.E., Heisterberg J.F. (1978) **Bat Records from Upper Volta, West Africa.** American Museum novitates, no. 2643.
- Kurt M.B. (1996) **Ausbreitung der Weißbrandfledermaus *Pipistrellus kuhlii* (KÜHL, 1819) in Österreich (Chiroptera, Vespertilionidae) Dem Andenken an Erich Kreissl.** Mitteilungen der Abteilung für Zoologie am Landesmuseum Joanneum Graz 50:17-24.
- Largen M.J., Kock D., Yalden D.W. (1974) **Catalogue of the mammals of Ethiopia. 1. Chiroptera.** Monitore Zoologico Italiano. Supplemento 5(1):221-298.
- Lavrenchenko L.A., Kruskop S.V., Bekele A., Belay G., Morozov P.N., Ivlev Y.F., Warshavsky A.A. (2010) **Mammals of the Babbile Elephant Sanctuary (Eastern Ethiopia).** Russina Journal of Theriology 9(2):47-60.
- Lavrenchenko L.A., Kruskop, S.V., Morozov, P.N. (2001) **Notes on the bats (Chiroptera) collected by the joint Ethiopian-Russian biological expedition, with remarks on their systematics, distribution, and ecology.** Bonner Zoologische Beiträge 52:127-147.
- Lino A., Fonseca C., Goiti U., Pereira M.J.R. (2014) **Prey Selection by *Rhinolophus hipposideros* (Chiroptera, Rhinolophidae) in a Modified Forest in Southwest Europe.** Acta Chiropterologica 16(1):75-83.
- Lučan R.K., Šálek M. (2013) **Observation of successful mobbing of an insectivorous bat, *Taphozous nudiventris* (Emballonuridae), on an avian predator, *Tyto alba* (Tytonidae).** Mammalia 77(2):235-236.
- Macy R.W., Heyneman D., Kuntz R.E. (1961) **Records of trematodes of the families Lecithodendriidae, Dicrocoeliidae, and Heterophyidae from Chiroptera collected in Egypt and Yemen, S. W. Arabia.** Proc. Helminthol. Soc. Washington 28:13-17.
- Madhavan A. (2010) **A catalogue of bats recorded in Thrissur district, kerala.** Zoo's Print Journal 15(11):355-358.
- Mahmood-ul-Hassan M., Javid A., Nadeem M.S., Ashraf S. (2012) **An extralimital record of the Egyptian tomb bat *Taphozous perforatus* from Pakistan.** Mammalia 76(2):227-229.
- Marinkelle C.J. (1977) **Trypanosoma (Herpetosoma) longiflagellum Sp.N. from the tomb bat, *Taphozous nudiventris*, from Iraq.** Journal of Wildlife Diseases, 13(3):262-264.
- McLellan L.J. (1986) **Notes on Bats of Sudan.** American Museum novitates, no. 2839.
- Mohammad K.M., Al-Zubaidi A.A.(2014) **Potentials of geodiversity for biodiversity at Ga'ara Depression, Iraqi Western Desert.** Advanced in Bioresearch 5(1): 169-177.

- Monadjem A., Schoeman M.C., Reside A., Pio D.V., Stoffberg S., Bayliss J., Cotterill F.P.D., Curran M., Kopp M., Taylor P.J. (2010) **A recent inventory of the bats of Mozambique with documentation of seven new species for the country.** Acta Chiropterologica 12(2):371-391.
- Motte G., Libois R. (2002) **Conservation of the lesser horseshoe bat (*Rhinolophus hipposideros* Bechstein, 1800) (Mammalia: Chiroptera) in Belgium. A case study of feeding habitat requirements.** Belgian Journal of Zoology 132(1):49-54.
- Nader I.A. (1975) **On the bats (Chiroptera) of the kingdom of Saudi Arabia.** Journal of Zoology 176(3):331-340.
- Nader I.A. (1982) **New distributional records of bats from the Kingdom of Saudi Arabia (Mammalia: Chiroptera).** Journal of Zoology 198(1):69-82.
- Nader I.A., Kock D. (1990) ***Eptesicus (Eptesicus) bottae* (Peters 1869) in Saudi Arabia with notes on its subspecies and distribution (Mammalia: Chiroptera: Vespertilionidae).** Senckenbergiana biol.70:1-13.
- Obuch J., Khaleghizadeh A. (2011) **Spatial Variation in the Diet of the Barn Owl *Tyto alba* in Iran.** Podoces 6(2):103-116.
- Osborn D.J. (1963) **New distributional records of bats from Turkey.** Mammalia 27(2):210-217.
- Osborn D.J. (1988) **New bat records from the Red Sea Mountains of Egypt.** Mammalia 52(4):596-598.
- Owen R.D., Qumsiyeh M.B. (1987) **The subspecies problem in the Trident leaf-nosed bat, *Asellia tridens*: homomorphism in widely separated populations.** Zeitschrift Für Säugetierkunde 6:329-337.
- Pearch M.J., Bates P.J.J., Magin C. (2001) **A review of the small mammal fauna of Djibouti and the results of a recent survey.** Mammalia 65(3):387-410.
- Pio D. (2010) **Evolutionary history and its relevance in understanding and conserving southern African biodiversity.** Ph.D. thesis, Université de Lausanne, Faculté de biologie et médecine.
- Popczyk B., Lesiński G., Baumann A., Wojtowicz B. (2008) **Kuhl's pipistrelle, *Pipistrellus kuhlii* (Kuhl, 1817) or *Pipistrellus lepidus* Blyth, 1845, in Central Poland - accidental record or a result of expansion?** Nyctalus (N.F.). Berlin 13(4):279-281.
- Puechmaille S., Hizem W., Allegrini B., Abiad A. (2012) **Bat fauna of Tunisia: review of records and new records, morphometrics and echolocation data.** Vespertilio 16:211-239.
- Purohit A., Senacha K.R. (2004) **Distribution of bats in and around Jaisalmer of the Great Indian Desert, India.** Vespertilio, 8:99-104.
- Purohit A., Somi P., Kaur A., Ram H. (2012) **First record of *Taphozous melanopogon* in the Barmer area of the Thar Desert, Rajasthan, India.** Vespertilio 16: 241-242.
- Qumsiyeh M.B. (1985) **The bats of Egypt.** Special Publication of the Texas Techological University, 23: 1-102.
- Qumsiyeh M.B. (1996) **Mammals of the Holy Land.** Texas Tech University Press, Lubbock.
- Qumsiyeh M.B., Amr Z.S., Al-Oran R.M. (1998) **Further records of bats from Jordan and a synopsis.** Turkish Journal of Zoology 22:277-284.
- Rautenbach I.L., Bronner G.N., Schlitter D.A.(1993) **Karyotypic data and attendant systematic implications for the bats of southern Africa.** Koedoe 36(2):87-104.
- Reiter A., Benda P., Hotový J. (2007) **First record of the Kuhl's Pipistrelle, *Pipistrellus kuhlii* (Kuhl, 1817), in the Czech Republic.** Lynx (N.S.), 38:47-54.
- Reiter G. (2004) **Postnatal growth and reproductive biology of *Rhinolophus hipposideros* (Chiroptera: Rhinolophidae).** Journal of Zoology 262(3):231-241.
- Reiter G., Pölzer E., Mixanig H., Bontadina F., Hüttmeir U. (2013) **Impact of landscape fragmentation on a specialised woodland bat, *Rhinolophus hipposideros*.** Mammalian Biology: Zeitschrift für Säugetierkunde 78(4):283-28.
- Robbins C.B. (1980) **Small mammals of Togo and Benin. I. Chiroptera.** Mammalia 44(1):83-88.
- Robbins L.W., Sarich V.M. (1988) **Evolutionary relationships in the family Emballonuridae (Chiroptera).** Journal of Mammalogy 69(1):1-13.

- Ruedas L.A., Lee T.E., Bickham J.W., Schlitter D.A. (1990) **Chromosomes of five species of Vespertilionid bats from Africa.** Journal of Mammalogy 71(1):94-100.
- Russo D., Jones G. (1999) **The social calls of Kuhl's pipistrelles *Pipistrellus kuhlii* (Kuhl, 1819): structure and variation (Chiroptera Vespertilionidae).** Journal of Zoology 249:4(476-481).
- Sachanowicz K., Bogdanowick W., Michalak S. (1999) **First record of *Taphozous nudiventris* Cretzschmar, 1830 (Chiroptera, Emballonuridae) in Turkey.** Mammalia 63(1):105–124.
- Sachanowicz K., Wower A., Bashta A.T. (2006) **Further range extension of *Pipistrellus kuhlii* (Kuhl, 1817) in central and eastern Europe.** Acta Chiropterologica, 8:543–548.
- Senacha K.R., Vyas K.B., Purohit A. (2006) **New records of short-nosed fruit bat *Cynopterus sphinx* (Vahl, 1797) from Thar desert, Rajasthan.** Zoo's Print Journal 21(10):2419-2420.
- Shah T.A., Srinivasulu C., Kaur H., Srinivasulu B., Devender G. (2014) **New distribution records of *Tadarida aegyptiaca* E. Geoffroy, 1818 (Mammalia: Chiroptera: Molossidae) from Karnataka, India.** International Journal of Fauna and Biological Studies 1(5):41-43.
- Shehab A., Daoud A., Kock D., Amr Z. (2004) **Small mammals recovered from owl pellets from Syria (Mammalia: Chiroptera, Rodentia).** Zoology in the Middle East, 33(1):27-42.
- Shehab A., Karataş A., Amr Z., Mamkhair I., Sözen M. (2007) **The distribution of bats (Mammalia: Chiroptera) in Syria.** Vertebrate Zoology 57:103–132.
- Shehab A., Mamkhair I., Amr Z. (2006) **First record of the Lesser Horseshoe bat, *Rhinolophus hipposideros* (Bechstein, 1800) (Rhinolophidae, Chiroptera) from Syria.** Hystrix, the Italian Journal of Mammalogy 17(2):161-166.
- Shehab A., Mamkhair I.H. (2004) **First record of the Egyptian Fruit Bat, *Rousettus aegyptiacus*, from Syria.** Zoology in the Middle East 33(1):73–78.
- Sinha Y.P. (1970) **Taxonomic notes on some Indian bats.** Mammalia 34(1):81-92.
- Sinha Y.P. (1976) **Bacula of Rajasthan bats.** Mammalia 40(1):97-104.
- Srinivasulu B., Srinivasulu C. (2007) **First specimen based record of the Egyptian free-tailed bat *Tadarida aegyptiaca* E. Geoffroy 1818 (Chiroptera: Molossidae) from Andhra Pradesh, India.** Zoo's Print Journal 22(12):2943.
- Srinivasulu C., Nagulu V. (2002) **Mammalian and avian diversity of the Nallamala Hills, Andhra Pradesh.** Zoo's Print Journal 17(1):675-684.
- Srinivasulu C., Srinivasulu B., Sinha Y.P. (2013) **Chiropteran fauna of Rajasthan: Taxonomy, distribution and status.** In: Sharma *et al.* (ed.) Faunal Heritage of Rajasthan, India: General Background and Ecology of Vertebrates.
- Stanley W.T., Goodman S.M. (2011) **Small mammal inventories in the east and west Usambara and South Pare Mountains, Tanzania. 3. Chiroptera.** Fieldiana Life and Earth Sciences Number 4:34-52.
- Thomas N.M., Harrison D.J., Bates P.J.J. (1994) **A study of the baculum in the genus *Nycteris* (Mammalia, Chroptera, Nycteridae) with consideration of its taxonomic importance.** Bonner Zoologische Beiträge 45:17-31.
- Thorn E., Peterhans J.K. (2009) **Small mammals of Uganda: Bats, shrews, hedgehog, golden-moles, otter-tenrec, elephant-shrews, and hares.** Zoologisches Forschungsmuseum Alexander Koenig, Bonn.
- Tourenq C., Khassim A., Sawaf M., Shuriqi M.K., Smart E., Ziolkowski M., Brook M., Selwan R., Perry L. (2009) **Characterisation of the Wadi Wurayah catchment basin, the First mountain Protected Area in the United Arab Emirates.** International Journal of Ecology and Environmental Sciences 35(4):289-311.
- Tourenq, C. and Drew, C.R. (2005) **The red list of terrestrial mammalian species of the Abu Dhabi Emirate.** Research report 03-31-0013-05, TERC Environmental Research and Wildlife Development Agency, Abu Dhabi, UAE.
- Trentin L., Rovero F. (2011) **Results of an inventory of bats in the Uzungwa scarp forest reserve, Tanzania.** Journal of East African Natural History 100(1-2):45-57.

- Van Cakenberghe V., De Vree F. (1998) **Systematics of African Nycteris (Mammalia: Chiroptera) Part III. The Nycteris thebaica group.** Bonner Zoologische Beitrage 48(2):123-166.
- Wassif K. (1954) **On a collection of mammals from northern Sinai.** Bulletin de l'Institute du Désert de l'Egypte 3(1):107-118.
- Wassif K., Madkour G., Soliman S. (1984) **On a collection of bats from Egypt.** Academy of Scientific Research & Technology, Fauna and Flora of Egypt, Cairo 2:1-36.
- Wawrocka K., Bartonička T., Reiter A. (2012) **Pipistrellus kuhlii, a bat species breeding and hibernating in the Czech Republic.** Vespertilio 16: 351-356.
- Webala P., Carugati C., Canova L., Fasola M. (2009) **Bat assemblages from eastern lake Turkana, Kenya.** Revue d'Ecologie (La Terre et la Vie) 64 (1):85-91.
- Williams C., Salter L., Jones G. (2011) **The winter diet of the lesser Horseshoe bat (*Rhinolophus hipposideros*) in Britain and Ireland.** Hystrix 22(1):159-166.
- Wolton R.J., Arak P.A., Godfray H.C.J., Wilson R.P. (1982) **Ecological and behavioural studies of the Megachiroptera at Mount Nimba, Liberia, with notes on Microchiroptera.** Mammalia 46(4):419-448.
- Yalden D.W, Largen M.J., Kock D., Hillman J.C. (1996) **Catalogue of the mammals of Ethiopia and Eritrea. 7. Revised checklist, zoogeography and conservation.** Tropical Zoology 9(1):73-164.
- Yassen A.E., Hassan H.A., Kawashti L.S. (1994) **Comparative study of the karyotypes of two Egyptian species of bats, *Taphozous perforatus* and *Taphozous nudiventris* (Chiroptera: Mammalia).** Experientia 50(11):1111-1114.
- Yom-Tov Y., Cadmon R. (1998) **Analysis of the distribution of insectivorous bats in Israel.** Diversity and Distributions 4(2):63-70.

## Appendix 3: Variables selection

List of initial covariates investigated for correlation/multi-collinearity and transformation applied. Final list used to run the model is marked in grey.

Variable		Transfo- rmation	VIF < 10	cor   < 0.7	GVIF < 3
Alt <sup>a</sup>	Altitude		yes	yes	yes
Bio1 <sup>a</sup>	Annual mean temperature	<sup>a</sup> 2			
Bio2 <sup>a</sup>	Mean diurnal range (mean of monthly (max temp - min temp))		yes	yes	yes
Bio3 <sup>a</sup>	Isothermality (Bio2/Bio7) (* 100)				
Bio4 <sup>a</sup>	Temperature seasonality (standard deviation *100)		yes	yes	yes
Bio5 <sup>a</sup>	Maximum temperature of warmest month				
Bio6 <sup>a</sup>	Minimum temperature of coldest month				
Bio7 <sup>a</sup>	Temperature annual range (Bio5-Bio6)				
Bio8 <sup>a</sup>	Mean temperature of wettest quarter	<sup>a</sup> 2	yes	yes	yes
Bio9 <sup>a</sup>	Mean temperature of driest quarter	<sup>a</sup> 2	yes	yes	yes
Bio10 <sup>a</sup>	Mean temperature of warmest quarter	<sup>a</sup> 2			
Bio11 <sup>a</sup>	Mean temperature of coldest quarter		yes	yes	
Bio12 <sup>a</sup>	Annual precipitation	sqrt			
Bio13 <sup>a</sup>	Precipitation of wettest month		yes		
Bio14 <sup>a</sup>	Precipitation of driest month		yes	yes	yes
Bio15 <sup>a</sup>	Precipitation seasonality (coefficient of variation)		yes	yes	yes
Bio16 <sup>a</sup>	Precipitation of wettest quarter	sqrt			
Bio17 <sup>a</sup>	Precipitation of driest quarter				
Bio18 <sup>a</sup>	Precipitation of warmest quarter		yes	yes	
Bio19 <sup>a</sup>	Precipitation of coldest quarter	log	yes	yes	yes
PET <sup>b</sup>	Potential evapotranspiration				
AI <sup>b</sup>	Aridity index	sqrt			
AET <sup>c</sup>	Actual evapotranspiration	sqrt			
SWB <sup>c</sup>	Soil-water balance	sqrt			
NDVI_Max <sup>d</sup>	Maximum NDVI (Normalized Difference Vegetation Index)				
NDVI_Min <sup>d</sup>	Minimum NDVI		yes	yes	yes
NDVI_Mean <sup>d</sup>	Mean NDVI				

<b>Variable</b>	<b>Transfo- rmation</b>	<b>VIF &lt; 10</b>	<b>  cor   &lt; 0.7</b>	<b>GVIF &lt; 3</b>
<b>NDVI_Range<sup>d</sup></b>	Range NDVI (maximum – minimum)			
<b>NDVI_SD<sup>d</sup></b>	Standard deviation of NDVI		yes	yes
<b>EVI_Max<sup>d</sup></b>	Maximum EVI (Enhanced Vegetation Index)			
<b>EVI_Min<sup>d</sup></b>	Minimum EVI		yes	
<b>EVI_Mean<sup>d</sup></b>	Mean EVI		yes	
<b>EVI_Range<sup>d</sup></b>	Range EVI (maximum - minimum)		yes	
<b>EVI_SD<sup>d</sup></b>	Standard deviation of EVI			

<sup>a</sup> WorldClim<sup>1</sup> : WorldClim provides a global dataset for the elevation and 19 bio-climatic variables interpolated from global monthly temperature and precipitation recordings from weather stations (Hijmans *et al.* 2005). Tiles for the overall study area were downloaded at 30 arc-seconds resolution (~1 km near the equator; using the *raster* R-package), then projected to Mollweide equal-area projection at  $5 \times 5 \text{ km}^2$  resolution. The high resolution of 30 arc-seconds was preferred over the 2.5 arc minutes (~5 km near the equator) in order not to lose much information while re-projecting. Instead of interpolation (assigning a value for the new pixel equals to the mean of the nearest three points at the original projection), a different approach was employed. First, a template grid covering the study area at the equal-area projection ( $5 \times 5 \text{ km}^2$ ) was prepared. Then, pixels of the original variables (at 30 arc-seconds resolution) were converted into centroid points (at the original projection: WGS-1984), then projected into Mollweide projection, then these points were rasterized using the mean value (or other relevant function for some variables; e.g. maximum for Bio5) of the points that spatially fall within each cell of the template grid. The same approach was used to prepare all other covariates used in this study.

<sup>b</sup> Global Potential Evapotranspiration (Global-PET) & Global Aridity (Zomer *et al.* 2007, 2008)<sup>2</sup> : available at the global scale at a resolution of 1 km, and were prepared based on models that use data from WorldClim as inputs.

<sup>c</sup> Global Actual Evapotranspiration (Global-AET) & Global Soil-Water-Balance (Global-SWB) (Trabucco & Zomer 2010)<sup>3</sup> : available at the global scale at a resolution of 1 km, and are prepared based on WorldClim and Global-PET database as primary input.

<sup>1</sup> [www.worldclim.org/bioclim/](http://www.worldclim.org/bioclim/)

<sup>2</sup> [www.cgiar-csi.org/data/global-aridity-and-pet-database](http://www.cgiar-csi.org/data/global-aridity-and-pet-database)

<sup>3</sup> [www.cgiar-csi.org/data/global-high-resolution-soil-water-balance](http://www.cgiar-csi.org/data/global-high-resolution-soil-water-balance)

<sup>d</sup> NDVI (Normalized Difference Vegetation Index) & EVI (Enhanced Vegetation Index) (Didan, 2015)<sup>4</sup> : map tiles for the whole study area were downloaded (and merged) for the period from 18/2/2000 to 22/3/2015 each 16 days (MODIS product: MOD13A2 – resolution: 1 km) using the MODIS R package (Mattiuzzi, 2014). Summary maps (maximum, minimum, mean, range, and standard deviation) across the whole period (for NDVI & EVI) were produced at the original MODIS scale/projection using a python code in ‘ArcGIS’ as this was memory intensive to perform in raster package of R. Each of the summary layers were then projected into the equal-area projection at  $5 \times 5 \text{ km}^2$  resolution.

- Possible covariate transformations were investigated, trying to keep some degree of uniform distributions across the range of each covariate (following: Dormann 2011). For some covariates, no transformation was effective, and they were kept untransformed.
- Variable selection: Multicollinearity amongst the potential covariates (covering the total study area) was assessed, maintaining highest GVIF (Generalized Variance Inflation Factor)  $< 3$  (Zuur et al. 2009) [which is equivalent, in our case, to maintaining a highest absolute correlation coefficient between each pair of covariates  $< 0.6$  and highest VIF (Variance Inflation Factor)  $< 10$ ]. This was also checked for each species’ study area.
- Aridity-related covariates (actual/potential evapotranspiration, aridity-index, and soil-water-balance) were all excluded as they show high collinearity with other WorldClim covariates; unsurprisingly as they were derived from models that use WorldClim data as input.
- Out of tensummary vegetation-related covariates, two NDVI covariates were used further: the cumulative minimum and the standard deviation NDVI. They reflect indices of minimum biomass and variability of vegetation-cover across the study area, respectively.

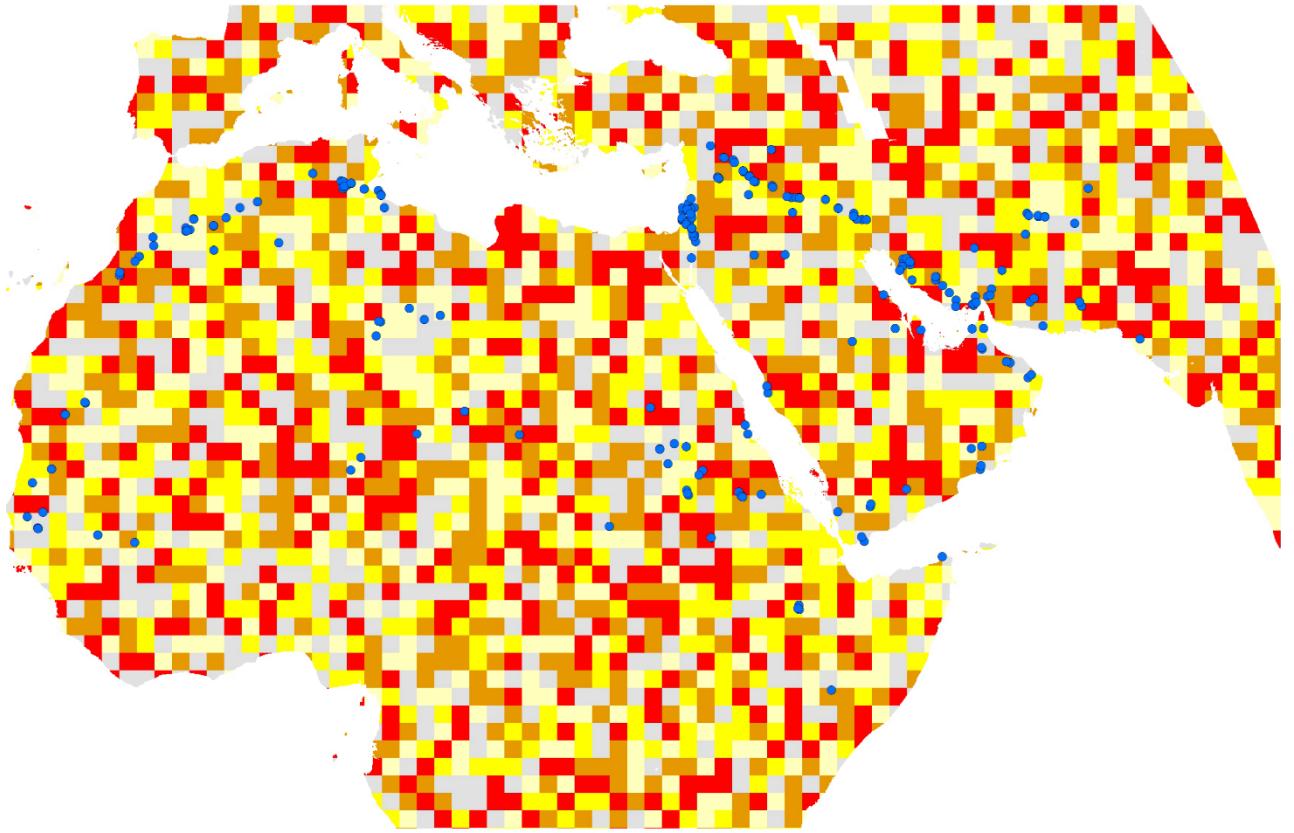
---

<sup>4</sup> [https://lpdaac.usgs.gov/dataset\\_discovery/modis/modis\\_products\\_table/mod13a2\\_v006](https://lpdaac.usgs.gov/dataset_discovery/modis/modis_products_table/mod13a2_v006)

## **Appendix 4: Per-species spatial-block cross-validation**

For each species, a different spatial-block cross-validation structure was used to run SDMs. Pixels across the study area were aggregated into larger blocks (each of  $20 \times 20$  cells =  $100 \times 100 \text{ km}^2$ ), and blocks were distributed into cross-validation folds. Presences and backgrounds in each block were used together either as training or testing (Fithian *et al.* 2015). The blocks were not distributed into cross-validation folds randomly, as this would have yielded highly unbalanced numbers of presence-locations across spatial folds. Instead, we balanced the number of presence locations at different folds by calculating their numbers at each block; the top 5 blocks were then, sequentially, randomly assigned to five folds (to avoid that the first fold always has the highest number of presences).

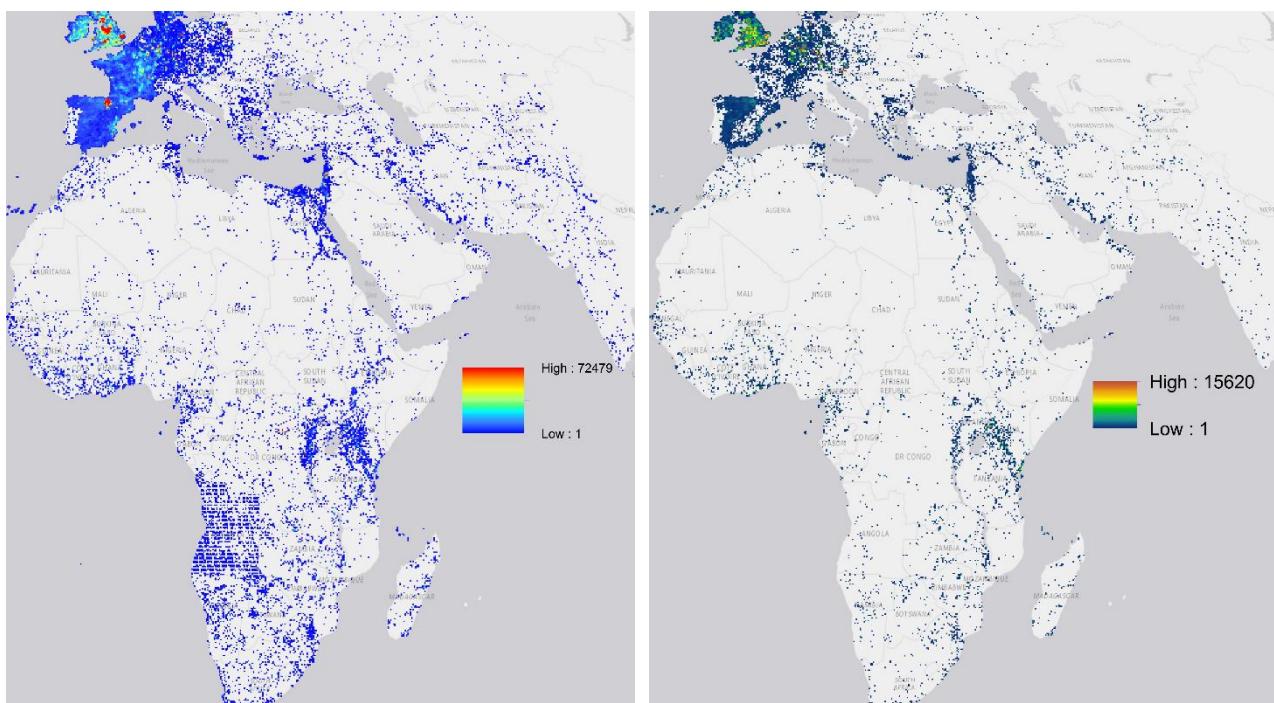
To avoid high variability in the environmental space between folds and minimise much extrapolation while predicting to the left-out-fold, blocks without any species records were distributed into folds depending on a mean index of their similarity to the overall study area: using the Multivariate Environmental Similarity Surfaces ‘MESS’. MESS was proposed by Elith *et al.* (2010) to identify areas of novel climates by comparing future (or past) climate to those used to run the models, as the interpretation of future projections at these locations should be considered with caution. We calculated the MESS-score for each pixel (and a mean value for each block) quite differently from how it was proposed in the first place: we measured the environmental similarity of each pixel to all available pixels in the study area (using the ‘dismo’ package). The original application of MESS gives most dissimilar locations more negative values (Elith *et al.* 2010); however here we expect no negative values as there is no climates novelty. Blocks without any presence locations are sorted in descending order (depending on their average MESS value), and then the highest five blocks were distributed randomly into different folds. This was repeated for all blocks until all blocks were distributed to different folds.



An example of how spatial-block cross-validation applied for *Asellia tridens*. Different colours represent how different blocks are distributed into cross-validation folds. Blue points represent the species known distribution from outside Egypt. For each per species' study area, a different blocking structure was prepared, to balance the number of presence locations and environmental variability at different cross-validation model runs.

## Appendix 5: Sampling-effort model

Data for closely related species (bats/non-marine mammals) were downloaded from GBIF (The Global Biodiversity Information Facility<sup>5</sup> – April 2015) and used (along with available focal species records: Tables 1 & A1) to model the relative sampling intensity (surveying effort) of bats/mammals. The prediction map of this model was used as bias covariate in the ‘Effort models’ (see main text, Fig. 1, bottom right). Records were assessed before usage (records with missing coordinates or with clear errors were excluded: e.g. missed latitude or longitude / equal latitude and longitude / records in the sea or the ocean / potentially swapped latitude and longitude), resulting in a total of 435,458 bat records / 2,039,158 non-marine mammal records (maps below). GBIF records (for both bats and mammals) show high bias towards the Western Europe compared to any other area in the study area, followed by scattered locations elsewhere (mostly close to the main cities, water bodies, populated areas, or seemingly a result of a mammals atlas mapping activity in southwest Africa). The majority of Africa and eastern Europe to western Asia is extremely under-represented in GBIF, with huge gaps in North Africa and Arabia.



Maps show the number of GBIF records of non-marine mammals (left) and bats (right) per a grid of  $20 \times 20 \text{ km}^2$  (accessed date: April 2015).

Initial trials for modelling the sampling effort were done using DWPR-GLM (Down-Weighted Poisson Regression) and Maxent (both using PPM-like approach; see main text); without much difference in the resulted prediction pattern (although Maxent model ‘using default feature classes and regularization multiplier’ shows, visually, an over-fitted spatial pattern). DWPR-GLM model

was chosen to run at  $5 \times 5$  km<sup>2</sup> equal-area projection: all the bats/mammals presence locations as the response (without duplicates removal) and using different covariate set than the environmental variables used to run the species SDMs (Merow et al. 2016):

- *Terrain roughness* (represented here as a per-pixel standard deviation of altitude): Altitude maps from WorldClim<sup>6</sup> was downloaded at of 30 arc-seconds resolution [ $\sim 1$  km near the equator], then projected as points in Mollwide equal-area projection. The points were then converted to a raster (rasterized), representing the standard deviation of the elevation values located at any target pixel at the resolution of  $5 \times 5$  km<sup>2</sup>;
- *Distance to main cities*<sup>7</sup> : the Euclidean distance between the centroid of each pixel and the nearest human settlement;
- *Distance to main roads*<sup>8</sup> : the Euclidean distance between the centroid of each pixel and the nearest road (Source: Global Roads Open Access Data Set ‘gROADS’);
- *Population count*<sup>9</sup> : global population count in 2000;
- Protection status<sup>10</sup> : a binary variable indicating the protection status of each pixel [source: The world Database of Protected Areas:].

The overall pattern of the predicted sampling effort for both bats and mammals were roughly similar (maps below), so the prediction from the bats sampling effort model was used further (as bias covariate in the effort model). It represents the overall relative abundance of bat species sightings per unit area (based only on non-climatic covariates).

---

<sup>5</sup> <http://www.gbif.org/>

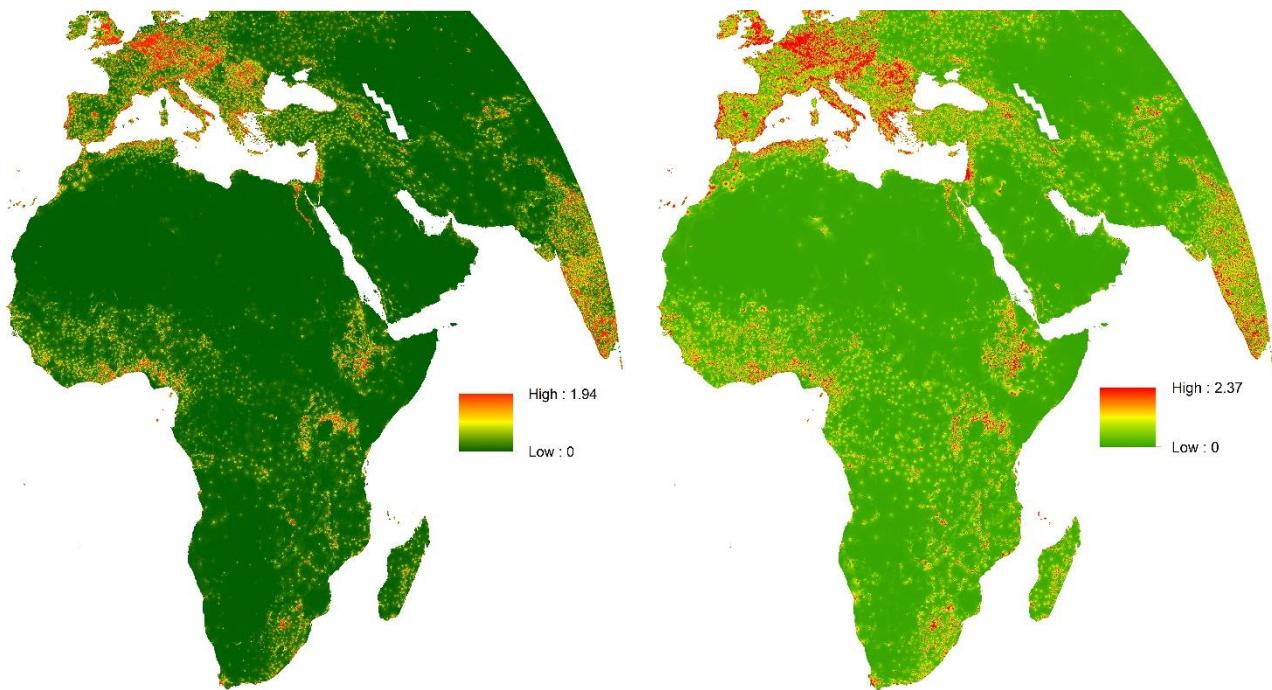
<sup>6</sup> <http://www.worldclim.org/>

<sup>7</sup> <http://sedac.ciesin.columbia.edu/data/set/grump-v1-settlement-points/data-download>; year: 2000

<sup>8</sup> <http://sedac.ciesin.columbia.edu/data/set/groads-global-roads-open-access-v1>

<sup>9</sup> <http://sedac.ciesin.columbia.edu/data/set/gpw-v3-population-count>;

<sup>10</sup> <http://protectedplanet.net>



The predicted sampling intensity of sightings (sampling effort) for all bats (left) and non-marine mammals (right) using the DWPR-GLM (Down-Weighted Poisson Regression) model.

## Appendix 6: Mixed effect model analysis of evaluation

Mixed Models formula:

**Evaluation metric ~ Modelling algorithms \* Bias models + total number of training presences (training data) + number of pixels in the study area (total area) + (1 | species)**

- Evaluation metric
  - continuous variable for either AUC or TSS
  - either using mean cross-validation evaluation ( $AUC_{cv}/TSS_{cv}$ ) or independent evaluation in Egypt (using mean prediction of 5-folds cross-validation to Egypt and independent testing data from Egypt)
- Modelling algorithms: categorical variable indicating the modelling algorithm used (elastic net / GLM / Maxent)
- Bias models:
  1. *Modelling evaluation*: evaluation of environment-only model (base line; no bias manipulation) is compared with those of bias-accounting models (accessibility and effort), without correction for sampling bias [without fixing bias variables at any values].
  2. *Bias correction evaluation*: evaluation of environment-only model (base line; no bias manipulation) is compared with those of bias accounting models (accessibility and effort), after correction for sampling bias [accessibility bias variables are set to zero, while the effort bias variable is set to either zero or the maximum estimated effort of the target species' presence records; see below].
- Species: random factor; categorical variable for the species.

## 1. Modelling Evaluation – Without Bias correction

### Variance explained (sum of squares)

	AUC		TSS	
	Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
Bias models	3.8	44.3	11.4	66.3
Model techniques	1.2	33.7	5.1	58.2
Bias models * Model techniques	0.2	5.5	0.7	11.5
Total number of training presences	0.005	0.03	0.02	0.043
Study area	0.004	0.02	0.05	0.039

### Parameter estimates (Mixed models summary)

	AUC		TSS	
	Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
(Intercept) [env-only/elastic net]	0.88036	0.7377	0.6928	0.5340
Effort.elastic net	0.00684	- 0.0026	0.0086	- 0.0149
Accessibility.elastic net	0.03815	0.1227	0.0660	0.1499
Env-only.Maxent	- 0.01480	0.0361	- 0.0281	0.0400
Effort.Maxent	0.00596	0.1096	0.0057	0.1434
Accessibility.Maxent	0.02399	0.1500	0.0325	0.1894
Env-only.GLM	0.01329	- 0.0380	0.0198	- 0.0480
Effort.GLM	0.02205	- 0.0383	0.0395	- 0.0500
Accessibility.GLM	0.03916	0.0626	0.0709	0.0652
Total number of training presences	0.03274	0.0418	0.0664	0.0842
Study area	- 0.02970	- 0.0347	- 0.0933	- 0.0797

### Estimated differences in the least square means

		AUC		TSS	
		Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
<b>Modelling techniques</b>	elastic net – Maxent	0.010	- 0.0585	0.0215	- 0.0793
	elastic net – GLM	- 0.010	0.0446	- 0.0185	0.0559
	Maxent – GLM	- 0.020	0.1031	- 0.0401	0.1352
	Env-only – Effort	- 0.012	- 0.0235	- 0.0207	- 0.0288
	Env-only – Accessibility	- 0.034	- 0.1124	- 0.0592	- 0.1375
	Effort – Accessibility	- 0.022	- 0.0889	- 0.0385	- 0.1087

### Estimated differences in the least square means (interactions)

			AUC		TSS	
			Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
<b>Bias models</b>	<b>elastic net</b>	Env-only – Effort	– 0.0068	0.0026	– 0.0086	0.0149
		Env-only – Accessibility	– 0.0381	– 0.1227	– 0.0660	– 0.1499
		Effort – Accessibility	– 0.0313	– 0.1254	– 0.0574	– 0.1647
	<b>GLM</b>	Env-only – Effort	– 0.0088	0.0003	– 0.0198	0.0020
		Env-only – Accessibility	– 0.0259	– 0.1006	– 0.0511	– 0.1133
		Effort – Accessibility	– 0.0171	– 0.1009	– 0.0314	– 0.1153
	<b>Maxent</b>	Env-only – Effort	– 0.0207	– 0.0735	– 0.0338	– 0.1034
		Env-only – Accessibility	– 0.0388	– 0.1139	– 0.0606	– 0.1494
		Effort – Effort	– 0.0180	– 0.0404	– 0.0268	– 0.0460
<b>Modelling techniques</b>	<b>elastic net – Maxent</b>	elastic net – Maxent	0.0009	– 0.1123	0.0029	– 0.1583
		elastic net – GLM	– 0.0152	0.0356	– 0.0309	0.0352
		Maxent – GLM	– 0.0161	0.1479	– 0.0339	0.1934
	<b>Accessibility</b>	elastic net – Maxent	0.0142	– 0.0273	0.0336	– 0.0396
		elastic net – GLM	– 0.0010	0.0602	– 0.0049	0.0846
		Maxent – GLM	– 0.0152	0.0874	– 0.0384	0.1242
	<b>Env-only</b>	elastic net – Maxent	0.0148	– 0.0361	0.0281	– 0.0400
		elastic net – GLM	– 0.0133	0.0380	– 0.0198	0.0480
		Maxent – GLM	– 0.0281	0.0741	– 0.0479	0.0880

## 2. Bias correction evaluation – using bias-free predictions

After correcting for sampling bias, initial trials show that using either value for fixing the effort bias covariate produce very similar evaluations (however, Maxent models show quite lower AUCs using the maximum effort value of training presences than for fixing at zero; Fig. 3), so we limited further analyses to effort<sub>Max</sub>.

When predicting with fixed values for the bias-covariates, modelling algorithms were most important for explaining variability of cross-validations, followed by sampling-bias models and then their interaction (for evaluation in Egypt highest variability was explained by the sampling-bias models, followed by the modelling algorithm, and then their interaction). Again, the number of training presences and study area were much less important (and the sign of their effects resembles those of modelling evaluation). The accessibility model had lower AUC compared to the two other sampling-bias models in all comparisons (Fig. 3). On cross-validation, bias-accounting models had lower AUC-values compared to the environment-only model (very little difference between environment-only and effort model for GLM; Fig. 3a). For evaluations in Egypt, environment-only and effort model were hardly different (effort model had lower AUC for Maxent-; Fig. 3b).

**Variance explained (sum of squares)**

	AUC		TSS	
	Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
Bias models	3.5	17.3	9.20	29.2
Model techniques	5.2	5.1	14.40	8.0
Bias models * Model techniques	0.3	2.5	0.96	2.7
Total number of training presences	0.010	0.04	0.04	0.06
Study area	0.007	0.03	0.07	0.06

**Parameter estimates (Mixed models summary)**

	AUC		TSS	
	Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
(Intercept) [env-only/ elastic net]	0.8804	0.7377	0.6928	0.5340
Effort.elastic net	-0.0118	-0.0030	-0.0188	-0.0195
Accessibility.elastic net	-0.0313	-0.0701	-0.0556	-0.1025
Env-only.Maxent	-0.0148	0.0361	-0.0281	0.0400
Effort.Maxent	-0.0385	0.0002	-0.0708	-0.0048
Accessibility.Maxent	-0.0565	-0.0664	-0.0892	-0.0794
Env-only.GLM	0.0133	-0.0380	0.0198	-0.0480
Effort.GLM	0.0117	-0.0368	0.0191	-0.0462
Accessibility.GLM	-0.0127	-0.0742	-0.0246	-0.1015
Total number of training presences	0.0439	0.0557	0.0821	0.1028
Study area	-0.0385	-0.0546	-0.1095	-0.1045

### Estimated differences in the least square means

		AUC		TSS	
		Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
<b>Modelling techniques</b>	elastic net – Maxent	0.0222	– 0.0143	0.0379	– 0.0259
	elastic net – GLM	– 0.0184	0.0253	– 0.0296	0.0246
	Maxent – GLM	– 0.0407	0.0396	– 0.0675	0.0505
	Env-only – Effort	0.0124	0.0125	0.0207	0.0208
	Env-only – Accessibility	0.0330	0.0696	0.0537	0.0918
	Effort – Accessibility	0.0207	0.0570	0.0330	0.0709

### Estimated differences in the least square means (interactions)

			AUC		TSS	
			Mean cross-validation	Egypt evaluation	Mean cross-validation	Egypt evaluation
<b>Bias models</b>	<b>elastic net</b>	Env-only – Effort	0.0118	0.0030	0.0188	0.0195
		Env-only – Accessibility	0.0313	0.0701	0.0556	0.1025
		Effort – Accessibility	0.0196	0.0671	0.0368	0.0830
	<b>GLM</b>	Env-only – Effort	0.0016	– 0.0012	0.0006	– 0.0018
		Env-only – Accessibility	0.0260	0.0362	0.0444	0.0535
		Effort – Accessibility	0.0244	0.0374	0.0437	0.0553
	<b>Maxent</b>	Env-only – Effort	0.0237	0.0359	0.0427	0.0448
		Env-only – Accessibility	0.0417	0.1025	0.0611	0.1194
		Effort – Accessibility	0.0180	0.0666	0.0184	0.0746
	<b>Effort</b>	elastic net – Maxent	0.0267	– 0.0032	0.0520	– 0.0146
		elastic net – GLM	– 0.0235	0.0338	– 0.0379	0.0267
		Maxent – GLM	– 0.0502	0.0370	– 0.0899	0.0414
	<b>Accessibility</b>	elastic net – Maxent	0.0252	– 0.0037	0.0336	– 0.0230
		elastic net – GLM	– 0.0186	0.0041	– 0.0310	– 0.0009
		Maxent – GLM	– 0.0437	0.0078	– 0.0646	0.0221
	<b>Env-only</b>	elastic net – Maxent	0.0148	– 0.0361	0.0281	– 0.0400
		elastic net – GLM	– 0.0133	0.0380	– 0.0198	0.0480
		Maxent – GLM	– 0.0281	0.0741	– 0.0479	0.0880

## Supplementary references

- Didan K (2015): MOD13A2 MODIS/Terra Vegetation Indices 16-Day L3 Global 1km SIN Grid V006. NASA EOSDIS Land Processes DAAC.
- Dormann CF (2011): Modelling Species' Distributions. In Fred J, Hauke R, Broder B (Eds.): Modelling Complex Ecological Dynamics. Berlin, Heidelberg: Springer Berlin Heidelberg, pp. 179–196.
- Elith J, Kearney M, Phillips S (2010): The art of modelling range-shifting species. Methods in Ecology and Evolution 1: 330–342.
- Fithian, W., Elith, J., Hastie, T., Keith, D.A. & O'Hara, R.B. (2015). Bias correction in species distribution models: pooling survey and collection data for multiple species. Methods in Ecology and Evolution, 6, 424–438.
- Friedman J, Hastie T, Tibshirani R (2010): Regularization Paths for Generalized Linear Models via Coordinate Descent. Journal of Statistical Software, 33(1), 1-22. URL: <http://www.jstatsoft.org/v33/i01/>.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A (2005): Very high resolution interpolated climate surfaces for global land areas. In Int. J. Climatol. 25 (15), pp. 1965–1978.
- Merow C, Allen JM, Aiello-Lammens M & Silander JA (2016): Improving niche and range estimates with Maxent and point process models by integrating spatially explicit information. Global Ecology and Biogeography, 25, 1022–1036.
- Mattiuzzi M (2014): MODIS: MODIS acquisition and processing. R package version 0.10-17/r484. (<https://R-Forge.R-project.org/projects/modis/>)
- Renner IW, Elith J, Baddeley A, Fithian W, Hastie T, Phillips SJ., *et al.* (2015): Point process models for presence-only analysis. Methods in Ecology and Evolution 6: 366–379.
- Trabucco A, Zomer RJ (2010): Global Soil Water Balance Geospatial Database. CGIAR Consortium for Spatial Information. Published online, available from the CGIAR-CSI GeoPortal at: <http://www.cgiar-csi.org>.
- Zomer RJ, Bossio DA, Trabucco A, Yuanjie L, Gupta DC, Singh VP (2007): Trees and Water: Smallholder Agroforestry on Irrigated Lands in Northern India. Colombo, Sri Lanka: International Water Management Institute. pp 45. (IWMI Research Report 122).
- Zomer RJ, Trabucco A, Bossio DA, van Straaten O, Verchot LV (2008): Climate Change Mitigation: A Spatial Analysis of Global Land Suitability for Clean Development Mechanism Afforestation and Reforestation. Agric. Ecosystems and Envir. 126: 67-80.
- Zuur AF, Ieno EN, Walker NJ, Saveliev AA, Smith GM (2009): Mixed Effects Models and Extensions in Ecology with R. New York, NY: Springer New York.